

PHENOLIC AND ANTIOXIDANT ACTIVITY IN ASIAN VEGETABLES

Vivek Saurabh

Division of Food Science and Postharvest Technology, ICAR – Indian Agricultural Research Institute, New Delhi–110012, India

Department of Food Science and Postharvest Technology, Bihar Agricultural University, Sabour Bhagalpur, Bihar–813210, India

Charanjit Kaur

Division of Food Science and Postharvest Technology, ICAR – Indian Agricultural Research Institute, New Delhi–110012, India

Manoj Kumar

Chemical and Biochemical Processing Division, ICAR – Central Institute for Research on Cotton Technology, Mumbai–400019, India

Keywords: Asian vegetables, Antioxidant, Functional food, Health, Phenols, Bioactivity, Bioavailability, Vegetable processing

Contents

1. Introduction
2. Classification of phenolic compounds
3. Antioxidant activity
4. Phenolics in Asian vegetables
 - 4.1. Beet Root
 - 4.2. Carrot
 - 4.3. Radish
 - 4.4. Sweet Potato
 - 4.5. Turmeric
 - 4.6. Spinach
 - 4.7. Fenugreek
 - 4.8. Amaranthus
 - 4.9. Chenopodium
 - 4.10. Coriander
 - 4.11. Asparagus
 - 4.12. Lettuce
 - 4.13. Fennel
 - 4.14. Mustard
 - 4.15. Cruciferous Vegetables
 - 4.16. Cowpea
 - 4.17. French Bean
 - 4.18. Broad Bean
 - 4.19. Brinjal
 - 4.20. Tomato
 - 4.21. Capsicum
 - 4.22. Onion and Shallot

- 4.23. Garlic
- 4.24. Cucurbitaceous Vegetables
- 4.25. Bamboo shoots
- 4.26. Jackfruit
- 4.27. Moringa
- 4.28. Asian Tuber Crops
- 4.29. Malabar spinach
- 5. Correlation between Phenolic Compounds and Antioxidant Activity
- 6. Effect of Processing and Storage on Phenolics in Vegetables
- 7. Bioavailability
- 8. Ways to Promote Vegetable Consumption in Children
- 9. Conclusions
- Acknowledgement
- Glossary
- Bibliography
- Biographical Sketches

Summary

Vegetables are rich source of dietary fiber, vitamins, phenolic compounds and minerals known to improve immunity and lower incidence of type-2 diabetes, cancer, and neurodegenerative disease. The present chapter aims to provide a comprehensive overview of the different classes of phenolics, their mechanism of action, bioavailability and associated antioxidant activity (AOX) present in foods. Detailed information on different phenolics present in different Asian vegetables is presented. The chapter also covers and summarizes the effect of various processing methods in improving the total phenolic content (TPC) and AOX of vegetables. Role of anthocyanin (ACN) rich vegetables and fermented vegetables in diet has also been highlighted. The information provided in the chapter will be of immense value to researchers, nutritionists, and students; for improving their diet by incorporating more healthy vegetables in diet.

1. Introduction

Fruits and vegetables (F&V) are an integral part of human nutrition as they are a source of vitamins, minerals and antioxidant essential for fulfilling our basic physiological needs. The World Health Organization (WHO) recommends consumption of at least 400 g of F&V each day or 5-a-day, to prevent non-communicable diseases such as heart disease, stroke and some types of cancer (Chawner & Hetherington, 2021). Overwhelming epidemiological evidence has demonstrated that consumption of F&V helps to counteract oxidative stress which is responsible for the pathogenesis of various lifestyle diseases such as cardiovascular, diabetes, Alzheimer's, arthritis, Parkinson's and cancer. The sedentary lifestyle, consumption of processed food, exposure to pollution and lack of exercise plays an important role in inducing oxidative stress in our body (Sharifi-Rad et al., 2020).

Oxidation is a normal necessary process that regulates the normal functioning of our body and generates free radicals. Free radicals are reactive oxygen species (ROS) that help fight pathogens and antigens; however, when they are produced in excessive

amounts, there is imbalance between ROS and defense related antioxidants. This situation leads to oxidative stress subsequently diseases. The most common ROS includes superoxide radical ($O^{\cdot-2}$), singlet oxygen (1O_2), hydrogen peroxide (H_2O_2), and hydroxyl radical (OH^{\cdot}). ROS damage lipid, protein, and DNA; break down the structural and cellular proteins. They also break lipid membrane and increase membrane fluidity and permeability. Protein damage involves site-specific amino acid modification, peptide chain fragmentation, cross-linked reaction product aggregation, enzymatic inactivation, and proteolysis. In DNA damage, they oxidize deoxyribose, breaks strand, modify bases and crosslink DNA-protein. High amounts of free radicals damage cells and cause apoptosis, leading to the pathogenesis of cancer, diabetes, and cardiovascular disease. Oxidative stress also stimulates the immune response and causes allergic diseases, such as asthma, allergic rhinitis, atopic dermatitis, or food allergies.

Antioxidants are compounds that are capable of quenching free radicals and break radical chain reactions, maintain haemostasis and prevent oxidative damage. A more biologically relevant definition of antioxidant is “a synthetic or natural substance added to products to prevent or delay their deterioration by the action of oxygen in the air (oxidation). In biochemical terms, antioxidants are enzymes or organic substances that are capable of counteracting oxidative stress in animal tissues. On the nutritional perspective, antioxidants can be classified into endogenous and exogenous antioxidants. Endogenous antioxidants are synthesized in the cells and include all enzymatic antioxidants along with some non-enzymatic ones (i.e., thiols antioxidants and coenzyme Q10). Whereas, exogenous antioxidants have to be supplemented through diet since their synthetic pathways are usually present only in microbial or plant cells. Common exogenous antioxidants include ascorbic acid, phenolics, vitamin E, β -carotene, organosulfur compounds, glucosinolates and selenium. Vegetables are surplus in antioxidants and have been conferred the status of ‘functional foods’ as they provide health-promoting effects besides fulfilling basic physiological needs of the body (Kaur & Kapoor, 2001). Vegetables like, beetroot, tomato, broccoli, and garlic are good examples of functional foods.

The term "Asian Vegetables" encompasses a diverse array of plant species that are either indigenous to Asia or have become deeply integrated into Asian diets over centuries due to historical events like colonization, migration, and trade. Understanding the nativity of these vegetables is complex, as many now-ubiquitous vegetables, such as cruciferous varieties, and potatoes were introduced to Asia from other parts of the world. Conversely, vegetables like bamboo shoots, moringa, yam, taro, luffa, jackfruit, amaranth, Malabar spinach and several other vegetables have roots in Asian soil and remain staples in traditional culinary practices across the continent. These vegetables not only contribute to the nutritional and medicinal value of Asian diets but also reflect the rich agricultural heritage of the region. While modern cultivation practices often follow market demands and global health trends, many of these traditional vegetables continue to be grown using age-old methods. Various examples of the Asian vegetables considered in this chapter are given in subsections 4.1. to 4.29.

The human body utilizes natural, endogenous, and synthetic antioxidants as a defense against reactive species. Convincing evidence from the dietary patterns such as ‘Mediterranean diet’ (MD) and ‘Japanese diet’ (JD) supports the beneficial health

effects of consuming diet rich in fruits, vegetables, whole grain legumes, olive oil, polyphenols and vitamins (Siervo et al., 2021). The MD has been recognized as the healthiest lifestyle by UNESCO and the Council of the EU; and also recommended by the US Department of Health (Maggi et al., 2021). Data from multiple randomized clinical trials have demonstrated the powerful effect of MD against oxidative stress, neurodegenerative diseases, inflammation, cardiovascular disease and type 2 diabetes. Japanese population also has been known for having the longest life expectancy in the world, with food being the key factor associated with the longevity of the population. Balanced consumption of energy, grains, vegetables, fruits, meat, fish, eggs, soy products, dairy products, and alcoholic beverages have positively contributed to longevity and lower risk of cerebrovascular disease in Japan. Lower mortality rates associated with the Japanese diet have been documented by 'Dietary Intake Reference Guide' (Htun et al., 2017). Overwhelming scientific evidence supports the role of vegetables-based diet and adherence to a JD and MD in improving cognitive decline, dementia and Alzheimer's disease.

There are specific benefits of consuming vegetables compared to fruits and selective promotion of vegetables may improve health and reduce premature mortality. They are low in fat, high in mineral and dietary fiber content which improves gut health. In addition, bioactive phenolics, protect health through various antioxidant mechanisms; inhibit the oxidation of low density cholesterol (LDL), platelet aggregation, and prevent carcinogen formation (Wallace et al., 2020).

2. Classification of Phenolic Compounds (PCs)

Phenolic compounds (PCs) are organic compounds characterized by a hydroxyl group (-OH) attached to a carbon atom of an aromatic ring. In simpler terms, phenol is a monohydroxy benzene (C_6H_5OH) and is also known as benzaldehyde, similar to alcohol but has a strong hydrogen bond (Figure 1). Based on the origin, PCs can be classified into natural and synthetic PCs.

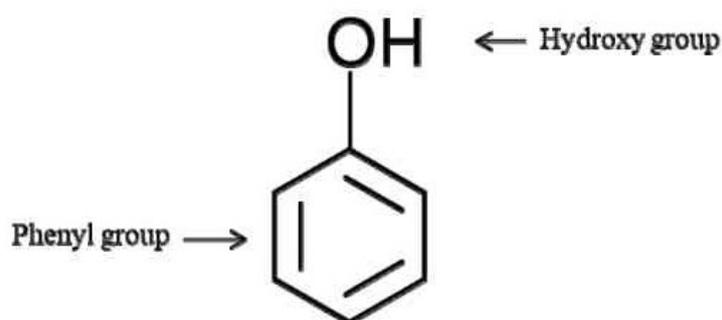


Figure 1. Basic structure of phenol

Natural PCs are the most abundant secondary metabolites and more than 8000 polyphenols have been identified in higher plants. These are widely found in plant-based foods (fruits, vegetables, cereals, olive, legumes, chocolate, etc.) and beverages (tea, coffee, beer, wine, etc.). PCs are responsible for sour, bitter and astringent taste of foods, for example, the bitter taste of spinach, lettuce, fenugreek and other vegetables.

Generally, they are produced naturally or in response to environmental stress, UV radiation and against insect pest attack. They are synthesized through two pathways; i) shikimic acid where phenylpropanoids are formed and ii) acetic acid pathway in which simple phenols are formed. In plants, PCs range from simple phenolic acids to highly polymerized compounds such as tannins. Common example of PCs includes ferulic acid, gallic acid, biphenols (resveratrol), and polyphenols (tannic acid). In plant cells, PCs occur in two forms namely, free or bound. The bound or insoluble form, attached to other molecules frequently to sugars (glycosyl residue) and proteins in cell walls of plant materials. Free and soluble PCs referred to as aglycones, that are less common, as they are toxic in the free state (Shahidi & Yeo, 2016). Most bound-insoluble phenolics are bound to cell wall substances such as fiber, pectin, cellulose and structural protein through covalent bonds and that constitute a relatively large amount (20-60%) in vegetables and legumes. Bound phenolics can be released by basic, acidic and enzymatic hydrolysis methods and show strong biological activities.

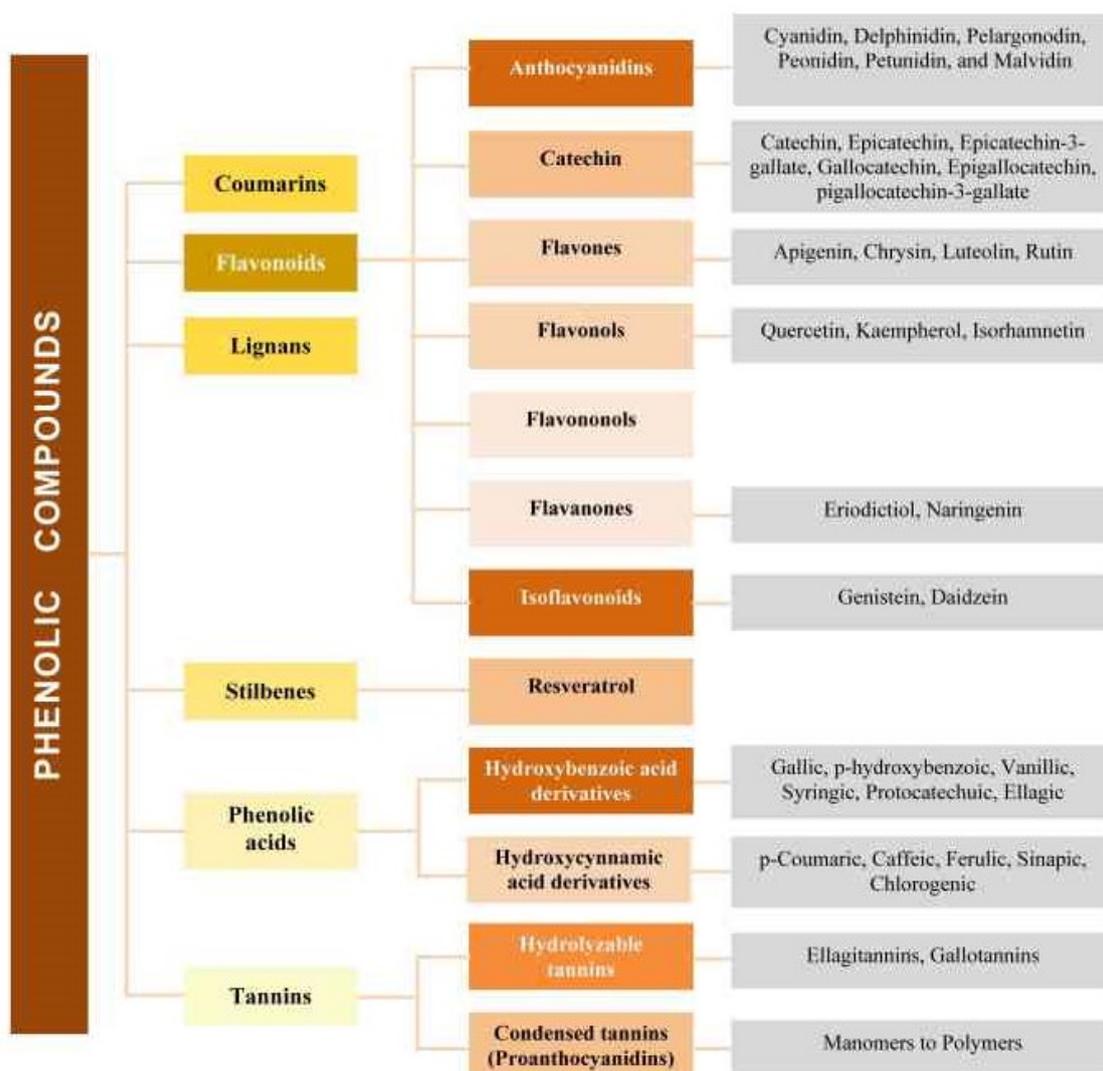


Figure 2. Classification of phenolic compound

Natural phenolics have been classified into six categories, viz. phenolic acid, flavonoids, stilbenes, coumarins, lignans and tannins. The major class of PCs is summarized in Figure 2. Substituted derivatives of hydroxybenzoic and hydroxycinnamic acids are the predominant phenolic acids in plants, later being the more common ones. Hydroxycinnamic acids include caffeic, p-coumaric and ferulic acids, which frequently occur in bound form as simple esters with quinic acid or glucose. A well-known bound hydroxycinnamic acid is chlorogenic acid, which is a combined caffeic and quinic acids. Hydroxybenzoic acid derivatives usually occur in the form of glucosides; p-hydroxybenzoic, vanillic and protocatechuic acids represent the most common forms (Shahidi & Ambigaipalan, 2015).



Figure 3a. Asian vegetables (Page1/3)

Flavonoids are the biggest subgroup, which is further divided on the basis of their structure into various subgroups like flavanols, flavanones, isoflavones, flavones and ACN. ACN are the major flavonoids responsible for blue, red, and purple coloration in

flowers and fruit. They have been referred to as ‘chameleon pigments’ as they change color depending upon the pH of cytoplasm. They appear as red under acidic conditions and blue under alkaline conditions. ACN occurs in the form of a glycoside of anthocyanidin (aglycone), common types of anthocyanidins present in vegetables are cyanidin, delphinidin, pelargonidin, peonidin, petunidin, and malvidin. Vegetables such as black carrot, red cabbage, red radish, red onions and purple potato are rich in ACN and possess high AOX (Figures 3a.b.c). Due to the broad spectrum of pharmacological properties of ACN, they are used for nutraceuticals in supplements. Tannins are high molecular (>500 Da) PCs generally referred to as anti-nutritional factors, as they have adverse effects on health. Tannins decrease protein digestibility and reduce the availability of essential amino acids. They are known to reduce the transport of thiamine and folic acid thereby decreasing their bioavailability. Polyphenols can also chelate iron and have inhibitory effects on the absorption of iron resulting in the deficiency of iron in the body.

Synthetic PCs are commonly used as preservatives in foods. These compounds are added to food to prevent or delay the onset of lipid oxidation during the processing and storage of foods. Currently permitted phenolics for food use include butylated hydroxyanisole (BHA, E-320), butylated hydroxytoluene (BHT, E-321), propyl gallate (PG, E-310) and tertiary-butylhydroquinone (TBHQ, E-319). Excessive addition of these synthetic phenolics has been reported to cause carcinogenicity, cytotoxicity, and oxidative stress. They can form molecular complexes with nucleic acid structure and cause damage to the double-helical structure of DNA. BHA is reported as the main cause of apoptosis in the human body. Toxic effects associated with their use, have prompted the food industry to look towards the natural phenolics in foods as preservative agents. As a result, a large number of phenolic rich plant extracts and by-products from the F&V processing industry are being explored for their efficacy as preservatives in lieu of synthetic ones.

-
-

TO ACCESS ALL THE 39 PAGES OF THIS CHAPTER,
Visit: <https://www.eolss.net/Eolss-sampleAllChapter.aspx>

Bibliography

References

- Alara, O. R., Abdurahman, N. H., & Ukaegbu, C. I. (2021). Extraction of phenolic compounds: A review. *Current Research in Food Science*, 4, 200–214. <https://doi.org/10.1016/j.crfs.2021.03.011>. [A comprehensive review on natural source of phenols and extraction of specific phenolic compound].
- Anantharaju, P. G., Gowda, P. C., Vimalambike, M. G., & Madhunapantula, S. V. (2016). An overview on the role of dietary phenolics for the treatment of cancers. *Nutrition Journal*, 15(1), 99. <https://doi.org/10.1186/s12937-016-0217-2>. [This study discusses about the role of plant phenol in modulation of genes related to initiation and progression of cancers].

Apak, R., Güçlü, K., Özyürek, M., & Karademir, S. E. (2004). Novel total antioxidant capacity index for dietary polyphenols and vitamins C and E, using their cupric ion reducing capability in the presence of neocuproine: CUPRAC method. *Journal of Agricultural and Food Chemistry*, 52(26), 7970–7981. <https://doi.org/10.1021/jf048741x>. [This presents a methodology for estimation of total antioxidant capacity using CUPRAC method].

Baliga, M. S., Shivashankara, A. R., Haniadka, R., Dsouza, J., & Bhat, H. P. (2011). Phytochemistry, nutritional and pharmacological properties of *Artocarpus heterophyllus* Lam (jackfruit): A review. *Food Research International*, 44(7), 1800-1811. [This review resent and discuss the phytochemistry, nutritional and pharmacological properties of jackfruit]

Benzie, I. F., & Strain, J. J. (1996). The ferric reducing ability of plasma (FRAP) as a measure of “antioxidant power”: the FRAP assay. *Analytical Biochemistry*, 239(1), 70–76. [A methodology to measure antioxidant power using FRAP assay].

Brand-Williams, W., Cuvelier, M. E., & Berset, C. (1995). Use of a free radical method to evaluate antioxidant activity. *LWT - Food Science and Technology*, 28(1), 25–30. [https://doi.org/10.1016/S0023-6438\(95\)80008-5](https://doi.org/10.1016/S0023-6438(95)80008-5). [This is a methodology for estimation of antioxidant activity using DPPH assay].

Bunea, A., Andjelkovic, M., Socaciu, C., Bobis, O., Neacsu, M., Verhé, R., & Camp, J. Van. (2008). Total and individual carotenoids and phenolic acids content in fresh, refrigerated and processed spinach (*Spinacia oleracea* L.). *Food Chemistry*, 108(2), 649–656. <https://doi.org/10.1016/j.foodchem.2007.11.056>. [A study on the effect of different processing and storage condition on the stability of carotenoid and phenolic acids of spinach].

Casquete, R., Velazquez, R., Hernandez, A., de Guia Cordoba, M., Aranda, E., Bartolome, T., & Martin, A. (2021). Evaluation of the quality and shelf-life of cayenne (*Capsicum spp.*). *LWT*, 145, 111338. <https://doi.org/10.1016/j.lwt.2021.111338>. [This study characterizes the quality parameters related to shelf life of different cayenne species].

Cecchi, L., Ieri, F., Vignolini, P., Mulinacci, N., & Romani, A. (2020). Characterization of volatile and flavonoid composition of different cuts of dried onion (*Allium cepa* L.) by HS-SPME-GC-MS, HS-SPME-GC×GC-TOF and HPLC-DAD. *Molecules*, 25(2), 408. <https://doi.org/10.3390/MOLECULES25020408>. [Characterized volatiles and flavonoids composition of dried onion of different cuts].

Chawner, L., & Hetherington, M. (2021). Utilising an integrated approach to developing liking for and consumption of vegetables in children. *Physiology & Behavior*, 238, 113493. <https://doi.org/10.1016/j.physbeh.2021.113493>. [A comprehensive discussion of factors involved in learning, integrated approach to enhance consumption of vegetables in children, practical application and gaps].

Cheng, D. M., Pogrebnyak, N., Kuhn, P., Poulev, A., Waterman, C., Rojas-Silva, P., Johnson, W. D., & Raskin, I. (2014). Polyphenol-rich Rutgers Scarlet Lettuce improves glucose metabolism and liver lipid accumulation in diet-induced obese C57BL/6 mice. *Nutrition*, 30(7–8), S52–S58. <https://doi.org/10.1016/j.nut.2014.02.022>. [This is a study on in-vitro and in-vivo antidiabetic potential of polyphenol-rich aqueous extract from Rutgers Scarlet Lettuce].

Cocksedge, S. P., Breese, B. C., Morgan, P. T., Nogueira, L., Thompson, C., Wylie, L. J., Jones, A. M., & Bailey, S. J. (2020). Influence of muscle oxygenation and nitrate-rich beetroot juice supplementation on O₂ uptake kinetics and exercise tolerance. *Nitric Oxide*, 99, 25–33. <https://doi.org/10.1016/j.niox.2020.03.007>. [This studies on the relation of beetroot juice on muscle oxygenation with exercise].

Dakhili, S., Abdolizadeh, L., Hosseini, S. M., Shojaee-Aliabadi, S., & Mirmoghtadaie, L. (2019). Quinoa protein: Composition, structure and functional properties. *Food Chemistry*, 299, 125161. <https://doi.org/10.1016/j.foodchem.2019.125161>.

Donma, M. M., & Donma, O. (2020). The effects of allium sativum on immunity within the scope of COVID-19 infection. *Medical Hypotheses*, 144, 109934. <https://doi.org/10.1016/j.mehy.2020.109934>. [A comprehensive review on functional properties, health promoting and immunity boosting characteristics of onion].

Duan, Y., Santiago, F. E. M., Dos Reis, A. R., de Figueiredo, M. A., Zhou, S., Thannhauser, T. W., & Li, L. (2021). Genotypic variation of flavonols and antioxidant capacity in broccoli. *Food Chemistry*, 338, 127997. <https://doi.org/10.1016/j.foodchem.2020.127997>. [This study the flavonoid and antioxidant activity of 15 accessions of broccoli].

Fleming, E., & Luo, Y. (2021). Co-delivery of synergistic antioxidants from food sources for the prevention of oxidative stress. *Journal of Agriculture and Food Research*, 3, 100107. <https://doi.org/10.1016/j.jafr.2021.100107>. [A comprehensive discussion on co-delivered bioactive compounds, their physicochemical interactions and the synergistic effects against oxidative stress].

Geberemeskel, G. A., Debebe, Y. G., & Nguse, N. A. (2019). Antidiabetic effect of fenugreek seed powder solution (*Trigonella foenum-graecum* L.) on hyperlipidemia in diabetic patients. *Journal of Diabetes Research*, 2019, 1–8. <https://doi.org/10.1155/2019/8507453>. [A case study on the Hyperlipidaemia in diabetics and effect of fenugreek seed powder solution].

Gonzali, S., & Perata, P. (2020). Anthocyanins from purple tomatoes as novel antioxidants to promote human health. *Antioxidants*, 9(10), 1017. <https://doi.org/10.3390/antiox9101017>. [This is a review on antioxidant potential of purple tomato anthocyanin and relation to human health].

Htun, N. C., Suga, H., Imai, S., Shimizu, W., & Takimoto, H. (2017). Food intake patterns and cardiovascular risk factors in Japanese adults: Analyses from the 2012 national health and nutrition survey, Japan. *Nutrition Journal*, 16(1), 61. <https://doi.org/10.1186/s12937-017-0284-z>. [A study on the role of Food intake patterns of Japan (Japanese diet) and cardiovascular risk factors].

Hussain, P. R., Suradkar, P., Javaid, S., Akram, H., & Parvez, S. (2016). Influence of postharvest gamma irradiation treatment on the content of bioactive compounds and antioxidant activity of fenugreek (*Trigonella foenum-graecum* L.) and spinach (*Spinacia oleracea* L.) leaves. *Innovative Food Science & Emerging Technologies*, 33, 268–281. <https://doi.org/10.1016/J.IFSET.2015.11.017>. [This studied the effect of bioactive compounds and antioxidant activity of fenugreek and spinach leaves].

Jayathilake, C., Visvanathan, R., Deen, A., Bangamuwage, R., Jayawardana, B. C., Nammi, S., & Liyanage, R. (2018). Cowpea: an overview on its nutritional facts and health benefits. *Journal of the Science of Food and Agriculture*, 98(13), 4793–4806. <https://doi.org/10.1002/JSFA.9074>.

Kahve, H. I., Akbulut, M., & Coklar, H. (2022). Identification and technological characterization of endogenous yeast isolated from fermented black carrot juice, shalgam. *LWT*, 154, 112823. <https://doi.org/10.1016/j.lwt.2021.112823>. [This study done for the identification of Pichia sp. to improve quality and safety of shalgam].

Kapusta-Duch, J., Szeląg-Sikora, A., Sikora, J., Niemiec, M., Gródek-Szostak, Z., Kuboń, M., Leszczyńska, T., & Borczak, B. (2019). Health-promoting properties of fresh and processed purple cauliflower. *Sustainability*, 11(15), 4008. <https://doi.org/10.3390/su11154008>.

Kaur, C., & Kapoor, H. C. (2001). Antioxidants in fruits and vegetables - The millennium's health. *International Journal of Food Science and Technology*, 36(7), 703–725. <https://doi.org/10.1046/j.1365-2621.2001.00513.x>. [A comprehensive review on the bioactive compounds of fruits and vegetables, their richness and estimation of antioxidant activity, correlation between intake and prevention of diseases].

Kaur, C., Nagal, S., Nishad, J., Kumar, R., & Sarika. (2014). Evaluating eggplant (*Solanum melongena* L.) genotypes for bioactive properties: A chemometric approach. *Food Research International*, 60, 205–211. <https://doi.org/10.1016/j.foodres.2013.09.049>. [Exploration of bioactive com compounds and antioxidant activity of eggplant genotype].

Kaur, C., Walia, S., Nagal, S., Walia, S., Singh, J., Singh, B. B., Saha, S., Singh, B., Kalia, P., Jaggi, S., & Sarika. (2013). Functional quality and antioxidant composition of selected tomato (*Solanum lycopersicon* L) cultivars grown in Northern India. *LWT - Food Science and Technology*, 50(1), 139–145. <https://doi.org/10.1016/j.lwt.2012.06.013>. [This study the functional quality and antioxidant composition of some tomato cultivars].

Khanam, U. K. S., Oba, S., Yanase, E., & Murakami, Y. (2012). Phenolic acids, flavonoids and total antioxidant capacity of selected leafy vegetables. *Journal of Functional Foods*, 4(4), 979–987. <https://doi.org/10.1016/j.jff.2012.07.006>. [A detailed study on bioactive compounds of some leafy vegetables].

Koley, T. K., Srivastava, S., Tripathi, Y. B., Banerjee, K., Oulkar, D., Goon, A., Tripathi, A., & Singh, B. (2019). High-resolution LCMS profiling of phenolic compounds of Indian black carrot and evaluation of its effect on antioxidant defense and glucose metabolism in animal model. *Agricultural Research*, 8(4), 481–489. <https://doi.org/10.1007/s40003-018-0389-4>. [A detailed analysis of Phenolic Compounds of Indian Black Carrot, Antioxidant Defense and Glucose Metabolism].

Kozioł, L., Knap, M., Sutor-Świeży, K., Górską, R., Dziedzic, E., Bieniasz, M., & Wybraniec, S. (2024). Identification and reactivity of pigments in prominent vegetable leaves of *Basella alba* L. var. 'Rubra' (Malabar spinach). *Food Chemistry*, 138714. [This is a study on pigment profiles of betacyanins from Malabar spinach]

Krochmal-Marczak, B., Zagórska-Dziok, M., Michalak, M., & Kiełtyka-Dadasiewicz, A. (2021). Comparative assessment of phenolic content, cellular antioxidant, antityrosinase and protective activities on skin cells of extracts from three sweet potato (*Ipomoea batatas* (L.) Lam.) cultivars. *Journal of King Saud University - Science*, 33(6), 101532. <https://doi.org/10.1016/j.jksus.2021.101532>. [This is a comparative study of bioactive compound and health benefiting activities of sweet potato extract].

Lee, H., Oh, I. N., Kim, J., Jung, D., Cuong, N. P., Kim, Y., Lee, J., Kwon, O., Park, S. U., Lim, Y., Kim, B., & Park, J. T. (2018). Phenolic compound profiles and their seasonal variations in new red-phenotype head-forming Chinese cabbages. *LWT*, 90, 433–439. <https://doi.org/10.1016/j.lwt.2017.12.056>. [This study analysed the deference in phenolic acid profile due to seasonal effect/variation].

Li, J., Huang, S. Y., Deng, Q., Li, G., Su, G., Liu, J., & David Wang, H. M. (2020). Extraction and characterization of phenolic compounds with antioxidant and antimicrobial activities from pickled radish. *Food and Chemical Toxicology*, 136, 111050. <https://doi.org/10.1016/j.fct.2019.111050>

Li, N., Wu, X., Zhuang, W., Xia, L., Chen, Y., Wu, C., Rao, Z., Du, L., Zhao, R., Yi, M., Wan, Q., & Zhou, Y. (2021). Tomato and lycopene and multiple health outcomes: Umbrella review. *Food Chemistry*, 343, 128396. <https://doi.org/10.1016/j.foodchem.2020.128396>. [A comprehensive review on the health benefits of lycopene and tomato].

Liang, Y., Li, Y., Zhang, L., & Liu, X. (2019). Phytochemicals and antioxidant activity in four varieties of head cabbages commonly consumed in China. *Food Production, Processing and Nutrition*, 1(1), 3. <https://doi.org/10.1186/s43014-019-0003-6>. [Study of bioactive compound present in some head cabbages of China and antioxidant activity].

Maggi, S., Rogoli, D., & Ecarnot, F. (2021). Healthy aging in the context of the Mediterranean diet–health–environment trilemma. *Aging and Health Research*, 1(2), 100015. <https://doi.org/10.1016/j.ahr.2021.100015>. [A detailed presentation on Mediterranean diet–health–environment trilemma].

Miller, N. J., Rice-Evans, C., Davies, M. J., Gopinathan, V., & Milner, A. (1993). A novel method for measuring antioxidant capacity and its application to monitoring the antioxidant status in premature neonates. *Clinical Science*, 84(4), 407–412. <https://doi.org/10.1042/cs0840407>. [This presents approaches to measure antioxidant power through absorbance of the ABTS*+ radical cation].

Mitharwal, S., Kumar, A., Chauhan, K., & Taneja, N. K. (2022). Nutritional, phytochemical composition and potential health benefits of taro (*Colocasia esculenta* L.) leaves: A review. *Food Chemistry*, 383, 132406. [A comprehensive study on composition and potential health benefits of taro].

Mizgier, P., Kucharska, A. Z., Sokół-Lętowska, A., Kolniak-Ostek, J., Kidoń, M., & Fecka, I. (2016). Characterization of phenolic compounds and antioxidant and anti-inflammatory properties of red cabbage and purple carrot extracts. *Journal of Functional Foods*, 21, 133–146. <https://doi.org/10.1016/j.jff.2015.12.004>.

Muñoz-Cuervo, I., Malapa, R., Michalet, S., Lebot, V., & Legendre, L. (2016). Secondary metabolite diversity in taro, *Colocasia esculenta* (L.) Schott, corms. *Journal of Food Composition and Analysis*, 52, 24–32.

Nandi, L. L., Saha, P., Behera, T. K., Lyngdoh, Y. A., Munshi, A. D., Saha, N. D., Hossain, F., Bhowmik, A., Pan, R. S., Verma, A., & Tomar, B. S. (2021). Genetic characterisation and population structure analysis of indigenous and exotic eggplant (*Solanum* spp.) accessions using microsatellite markers. *The Journal of Horticultural Science and Biotechnology*, 96(1), 73–86. <https://doi.org/10.1080/14620316.2020.1763211>.

- Oh, S., Tsukamoto, C., Kim, K., & Choi, M. (2017). Investigation of glucosinolates, and the antioxidant activity of Dolsan leaf mustard kimchi extract using HPLC and LC-PDA-MS/MS. *Journal of Food Biochemistry*, 41(3), e12366. <https://doi.org/10.1111/jfbc.12366>.
- Ou, B., Huang, D., Hampsch-Woodill, M., Flanagan, J. A., & Deemer, E. K. (2002). Analysis of antioxidant activities of common vegetables employing oxygen radical absorbance capacity (ORAC) and ferric reducing antioxidant power (FRAP) assays: A comparative study. *Journal of Agricultural and Food Chemistry*, 50(11), 3122–3128. <https://doi.org/10.1021/jf0116606>
- Pacifico, S., Galasso, S., Piccolella, S., Kretschmer, N., Pan, S. P., Nocera, P., Lettieri, A., Bauer, R., & Monaco, P. (2018). Winter wild fennel leaves as a source of anti-inflammatory and antioxidant polyphenols. *Arabian Journal of Chemistry*, 11(4), 513–524. <https://doi.org/10.1016/j.arabjc.2015.06.026>. [Study on the health benefitting properties of funnel leaves].
- Pires-Cabral, P., Pires-Cabral, P., & Quintas, C. (2021). *Salicornia ramosissima* as a salt substitute in the fermentation of white cabbage. *Journal of Food Science and Technology*. <https://doi.org/10.1007/s13197-021-05047-y>.
- Poveda, J., Zabalgoitia, I., Soengas, P., Rodríguez, V. M., Carrea, M. E., Abilleira, R., & Velasco, P. (2020). *Brassica oleracea* var. *acephala* (kale) improvement by biological activity of root endophytic fungi. *Scientific Reports*, 10(1), 20224. <https://doi.org/10.1038/s41598-020-77215-7>. [The diversity of root endophytes associated with different accessions of kale was studied].
- Prior, R. L., Wu, X., & Schaich, K. (2005). Standardized methods for the determination of antioxidant capacity and phenolics in foods and dietary supplements. *Journal of Agricultural and Food Chemistry*, 53(10), 4290–4302. <https://doi.org/10.1021/jf0502698>. [This paper provides an overview on basis and rationale for developing standardized antioxidant capacity methods for various industries].
- Rahate, K. A., Madhumita, M., & Prabhakar, P. K. (2021). Nutritional composition, anti-nutritional factors, pretreatments-cum-processing impact and food formulation potential of faba bean (*Vicia faba* L.): A comprehensive review. *LWT*, 138, 110796. <https://doi.org/10.1016/j.lwt.2020.110796>.
- Ríos-Ríos, K. L., Montilla, A., Olano, A., & Villamiel, M. (2019). Physicochemical changes and sensorial properties during black garlic elaboration: A review. *Trends in Food Science & Technology*, 88, 459–467. <https://doi.org/10.1016/j.tifs.2019.04.016>. [A comprehensive review on the changes occurs during black garlic preparation].
- Rolnik, A., & Olas, B. (2020). Vegetables from the Cucurbitaceae family and their products: Positive effect on human health. *Nutrition*, 78, 110788. <https://doi.org/10.1016/j.nut.2020.110788>. [This discusses about the bioactive compound present in cucurbitaceous crops and their role in human health].
- Salehi, B., Sharifi-Rad, J., Cappellini, F., Reiner, Ž., Zorzan, D., Imran, M., Sener, B., Kilic, M., El-Shazly, M., Fahmy, N. M., Al-Sayed, E., Martorell, M., Tonelli, C., Petroni, K., Docea, A. O., Calina, D., & Maroyi, A. (2020). The therapeutic potential of anthocyanins: current approaches based on their molecular mechanism of action. *Frontiers in Pharmacology*, 11. <https://doi.org/10.3389/fphar.2020.01300>. [A comprehensive review on class of anthocyanin, their bioavailability, therapeutic potential, molecular mechanisms of action, and their clinical significance].
- Satheesh, N., & Fanta, S. W. (2020). Kale: Review on nutritional composition, bio-active compounds, anti-nutritional factors, health beneficial properties and value-added products. *Cogent Food & Agriculture*, 6(1), 1811048. <https://doi.org/10.1080/23311932.2020.1811048>.
- Scalzo, R. L., Fibiani, M., Francese, G., D'Alessandro, A., Rotino, G. L., Conte, P., & Mennella, G. (2016). Cooking influence on physico-chemical fruit characteristics of eggplant (*Solanum melongena* L.). *Food Chemistry*, 194, 835–842. <https://doi.org/10.1016/j.foodchem.2015.08.063>. [A case study on the effect of cooking on physico-chemical characteristics of eggplant].
- Shahidi, F., & Ambigaipalan, P. (2015). Phenolics and polyphenolics in foods, beverages and spices: Antioxidant activity and health effects – A review. *Journal of Functional Foods*, 18, 820–897. <https://doi.org/10.1016/j.jff.2015.06.018>.
- Shahidi, F., & Yeo, J. (2016). Insoluble-bound phenolics in food. *Molecules*, 21(9), 1216. <https://doi.org/10.3390/molecules21091216>. [A comprehensive review on formation, localization, synthesis, and transfer insoluble-bound phenolics in plant].

Shahidi, F., & Zhong, Y. (2015). Measurement of antioxidant activity. *Journal of Functional Foods*, 18, 757–781. <https://doi.org/10.1016/j.jff.2015.01.047>. [This review describes the various methods available to estimate antioxidant activity of different oxidation substrates, with varying reaction mechanisms and conditions].

Sharifi-Rad, M., Kumar, N. V. A., Zucca, P., Varoni, E. M., Dini, L., Panzarini, E., Rajkovic, J., Fokou, P. V. T., Azzini, E., Peluso, I., Mishra, A. P., Nigam, M., El Rayess, Y., Beyrouthy, M. El, Polito, L., Iriti, M., Martins, N., Martorell, M., Docea, A., Setzer, W. N., Calina, D., Cho, W. C., & Sharifi-Rad, J. (2020). Lifestyle, oxidative stress, and antioxidants: Back and forth in the pathophysiology of chronic diseases. *Frontiers in Physiology*, 11, 694. <https://doi.org/10.3389/fphys.2020.00694>. [This review has discussed about the factors responsible for oxidative stress and increasing burden of chronic diseases, as well as the role of antioxidants in healthy ageing].

Siervo, M., Shannon, O. M., Llewellyn, D. J., Stephan, B. C., & Fontana, L. (2021). Mediterranean diet and cognitive function: From methodology to mechanisms of action. *Free Radical Biology and Medicine*, 176, 105–117. <https://doi.org/10.1016/j.freeradbiomed.2021.09.018>. [A study on Mediterranean diet and its beneficial role in human health].

Singleton, V. L., Orthofer, R., & Lamuela-Raventós, R. M. (1999). [14] Analysis of total phenols and other oxidation substrates and antioxidants by means of folin-ciocalteu reagent. In *Methods in Enzymology*, 299, 152–178. [https://doi.org/10.1016/S0076-6879\(99\)99017-1](https://doi.org/10.1016/S0076-6879(99)99017-1). [This is a methodology to estimate total phenolic content and antioxidant capacity using folin-ciocalteu reagent]

Tang, Y., & Tsao, R. (2017). Phytochemicals in quinoa and amaranth grains and their antioxidant, anti-inflammatory, and potential health beneficial effects: a review. *Molecular Nutrition & Food Research*, 61(7), 1600767. <https://doi.org/10.1002/mnfr.201600767>. [A comprehensive review on potential health benefitting effects of quinoa and amaranth grains].

Wallace, T. C., Bailey, R. L., Blumberg, J. B., Burton-Freeman, B., Chen, C. O., Crowe-White, K. M., Drewnowski, A., Hooshmand, S., Johnson, E., Lewis, R., Murray, R., Shapses, S. A., & Wang, D. D. (2020). Fruits, vegetables, and health: A comprehensive narrative, umbrella review of the science and recommendations for enhanced public policy to improve intake. *Critical Reviews in Food Science and Nutrition*, 60(13), 2174–2211. <https://doi.org/10.1080/10408398.2019.1632258>. [An umbrella review on the relation of fruits and vegetables with health and policy recommendations for increased consumption].

Wang, P., Shan, N., Ali, A., Sun, J., Luo, S., Xiao, Y., & Zhou, Q. (2022). Comprehensive evaluation of functional components, biological activities, and minerals of yam species (*Dioscorea polystachya* and *D. alata*) from China. *LWT*, 168, 113964. [Study of the species of yam for its functional components, biological activities, and minerals].

Wastyk, H. C., Fragiadakis, G. K., Perelman, D., Dahan, D., Merrill, B. D., Yu, F. B., Topf, M., Gonzalez, C. G., Van Treuren, W., Han, S., Robinson, J. L., Elias, J. E., Sonnenburg, E. D., Gardner, C. D., & Sonnenburg, J. L. (2021). Gut-microbiota-targeted diets modulate human immune status. *Cell*, 184(16), 4137-4153.e14. <https://doi.org/10.1016/j.cell.2021.06.019>. [This is a study on high fiber, fermented food, their interaction to human microbiome-immune system axis and reducing Non-communicable chronic diseases].

Wenli, S., Mohamad, H. S., & Qi, C. (2019). The insight and survey on medicinal properties and nutritive components of Shallot. *Journal of Medicinal Plants Research*, 13(18), 452–457. <https://doi.org/10.5897/JMPR2019.6836>. [This article describes about the phytochemicals present in shallot and their medicinal properties].

Wongmekiat, O., Leelarugrayub, N., & Thamprasert, K. (2008). Beneficial effect of shallot (*Allium ascalonicum* L.) extract on cyclosporine nephrotoxicity in rats. *Food and Chemical Toxicology*, 46(5), 1844–1850. <https://doi.org/10.1016/j.fct.2008.01.029>. [A study on beneficial effect of shallot extract on oxidant-induced disease nephrotoxicity].

Yadav, L. P., Gangadhara, K., Apparao, V. V., Singh, A. K., Rane, J., Kaushik, P., Sekhawat, N., Malhotra, S. K., Ramniwas., Rai, A. K., Yadav, S. L. & Berwal, M. K. (2024). Nutritional, antioxidants and protein profiling of leaves of *Moringa oleifera* germplasm. *South African Journal of Botany*, 165, 443–454. [A study on the chemical profiling of different germplasm of moringa].

Yang, Q. Q., Gan, R. Y., Ge, Y. Y., Zhang, D., & Corke, H. (2018). Polyphenols in common beans (*Phaseolus vulgaris* L.): Chemistry, analysis, and factors affecting composition. *Comprehensive Reviews in Food Science and Food Safety*, 17(6), 1518–1539. <https://doi.org/10.1111/1541-4337.12391>. [A comprehensive detail on the polyphenols in common beans and factors affecting their compositions].

Yang, M., Tao, L., Kang, X. R., Wang, Z. L., Su, L. Y., Li, L. F., Gu, F., Zhao, C. C., Sheng, J., Tian, Y. & Tian, Y. (2023). *Moringa oleifera* Lam. leaves as new raw food material: A review of its nutritional composition, functional properties, and comprehensive application. *Trends in Food Science & Technology*, 138, 399-416. [A review on nutritional composition, functional properties, and comprehensive application of moringa leaves].

Zhang, Y., Lv, C., Sun, J., Song, X., Makaza, N., & Wu, Y. (2021). Protective effects of broccoli extracts and sulforaphane against hydrogen peroxide induced oxidative stress in B16 cells. *Journal of Functional Foods*, 87, 104833. <https://doi.org/10.1016/j.jff.2021.104833>. [This study shows the protective effect of sulforaphane from broccoli extract on oxidative stress in B16 cells].

Biographical Sketches

Mr. Vivek Saurabh is an Assistant Professor-cum-Junior Scientist (Postharvest Technology) at Bihar Agricultural University, Sabour, India. His major field of interest is extraction of bioactive compounds namely pectin from waste and their application as edible coating for shelf-life extension of fruits and vegetables. He has published more than 25 research and review articles in high impact factor peer reviewed journals. He is an aspiring and a budding research scientist in the field of postharvest management.

Dr. Charanjit Kaur is a fellow of National Academy of Sciences (NASc), India and Professor in the Division of Food Science and Postharvest Technology. Her research interest is in the area of “Developing high value functional foods and nutraceutical ingredients from horticultural crops”. Her significant work envisages use of un-conventional processing through enzymes, and microwave and ultrasound assisted techniques for enhancing recovery of bio-active phyto-chemicals from waste. She has guided more than 20 MSc and PhD students and currently has 8 PhD students under her supervision. She has published nearly 130 research articles in high impact factor journal with total citations of 8013, h-index of 34 and i-10 index of 72. Her students have many acclaimed awards including ‘IARI Merit Gold Medal’, ‘Jawaharlal Nehru Award’, ‘Prime-Minister Fellowship’ and ‘DuPont innovation award’ to their credit.

Dr. Manoj Kumar is a Scientist (Plant Biochemistry) at ICAR–Central Institute for Research on Cotton Technology, Mumbai, India. He is a recipient of prestigious ‘Prime Minister Fellowship award’ and ‘IARI Gold medal’ for his outstanding research on natural colorants from black carrot. His research interest is in the field of extraction and characterization of proteins and other bioactive compounds from plant-based matrices. Currently he is on deputation to East Carolina University, Greenville, North Carolina, USA for undertaking training under CRISPR/Cas Technology funded by SERB, DST, India. He has published more than 150 research and review articles in peer-reviewed international journals.