

BIOCATALYTIC PROCESSES

Lucia Gardossi

Dipartimento di Scienze Farmaceutiche, Università degli Studi di Trieste, Trieste, Italy.

Francesco Molinari

Dipartimento di Scienze e Tecnologie Alimentari e Microbiologiche, Università degli Studi di Milano, Milano, Italy.

Keywords: Biocatalysis, enzyme, fermentation, microbiology, enantioselectivity, regioselectivity, chemoselectivity, protein engineering, immobilization, green chemistry.

Contents

1. Biocatalysis: definitions
 2. Traditional and modern applications of enzymes and biocatalysts
 3. Advantages and drawbacks related to the use of biocatalysts in chemistry
 4. Classification of enzymes used in organic synthesis.
 5. Enzymes most commonly employed in organic synthesis
 6. Biocatalysts in enantiotransformations
 7. Selecting and designing biocatalysts for practical applications
 8. Immobilization of biocatalysts
 9. Reaction media for biocatalysis
 10. Special techniques for the use of biocatalysts
 11. Industrial applications of biocatalysts: economic issues
 12. Examples of large scale productive processes employing biocatalysts in chemical and pharmaceutical industry
 13. Conclusions
- Acknowledgements
Glossary
Bibliography
Biographical Sketches

Summary

Enzymes form an abundant class of very effective and precise (bio)-catalysts that perform and regulate the processes in living matter. Biocatalysis exploits the ability of enzymes to transform also compounds that are not their natural substrates. Biocatalysis represents already an important tool for the production of fine chemicals and especially pharmaceuticals. Biocatalysts can be used as free or immobilized enzymes as well as microbial or plant cells.

The surge in practical utilization of biocatalysts is driven by their versatility, regio-, chemo-, enantioselectivity along with the necessity of chemical industry to translate to environmentally compatible catalysts and processes.

Novel methodologies for discovering industrial biocatalysts based on new techniques for microbial screening and molecular biology (including directed evolution and metabolic engineering) have led to the production of stable biocatalysts with customized activity and selectivity. Conversely, novel biocatalyst formulation based on innovative immobilization techniques has resulted in improved types of highly stable and efficient biocatalysts.

1. Biocatalysis: Definitions

1.1. Enzymes as Biocatalysts

Chemical transformations carried out by every living organism are enabled by thousands of proteins (enzymes) which have catalytic activity for conversion of a particular set of substrates to specific products. Biocatalysis is the general term for the transformation of natural and non-natural compounds by enzymes. Because of this, the term biocatalysis is also referred to the application of enzymes in chemistry (Bommarius and Riebel, 2004).

Over 3,000 enzymes have so far been identified, and this number will greatly increase thanks to the contribution of genomic and proteomic research. As catalysts, enzymes have remarkable specificities and sometimes phenomenal rate accelerations. A wide array of complex molecules is accepted by enzymes, including synthetic molecules with structures very different from the substrates found in nature. Biocatalysts are also endowed with selectivity, catalyzing reactions with unique chiral (stereo-) and positional (regio-) selectivities. The basis for the action of all enzymes as chemo-, regio- and stereospecific catalysts lies in their structure. Out of 20 amino acids, 19 of them are enantiopure L-amino acids providing an asymmetric microenvironment for substrate binding and subsequent chemical transformation in the enzyme active site.

These features make biocatalysis attractive as a complementary tool for transformations both in organic chemistry and in industry (Liese et al., 2006)

1.2. Parameters Affecting the Efficiency of Biocatalysts

Enzymes are able to accelerate the rate of some reactions by a factor of over 1000, as, for instance, in the protease-catalyzed hydrolysis of peptide bonds. Enzymes accelerate reactions by lowering the free energy of the transition state of a given reaction. Firstly the enzyme recognizes the substrate and forms an enzyme-substrate complex (ES), which is then converted into the product (P) (Figure 1).

Several parameters affect the practicality of an enzymatic reaction. Of particular importance are the specific activity (quantified by k_{cat}), specificity (determined by k_{cat}/K_M). In addition, the degree of inhibition by substrate or product (often determined by their affinity to the enzyme, K_M and K_P) may be particularly important in the outcome of a reaction. Ideally, the enzyme should have high specific activity and stability, and should undergo minimal substrate and product inhibition. Furthermore, the extent of substrate specificity can determine whether a given enzyme will have general

synthetic utility. Although enzymes with narrow substrate specificity are often efficient in catalyzing reactions using their natural substrate, this property becomes a limitation when developing catalysts for general purposes (Koeller and Wong, 2001).

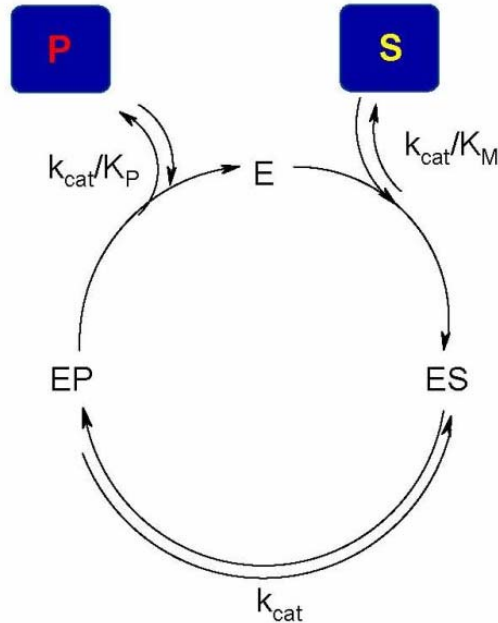


Figure 1. Parameters affecting enzymatic transformations

Finally, the stability of enzymes to environmental factors represents a further parameter of fundamental importance for the practical applications of biocatalysts.

2. Traditional and Modern Applications of Enzymes and Biocatalysts

Enzymes found in nature have been used since ancient times in the production of food products, such as cheese, sourdough, beer, wine and vinegar, and in the manufacture of commodities such as leather, indigo and linen. All of these processes relied on either enzymes produced by cells of spontaneously growing microorganisms or produced by plant and animal cells present in added preparations such as rumen of calf or papaya fruit. The development of fermentation processes and biochemical methods specifically aimed at the production of enzymes made it possible to manufacture enzymes as purified, well-characterized preparations even on a large scale. This development allowed the introduction of enzymes into true industrial products and processes, for example in the formulation of detergents and in textile and starch industries. The use of recombinant gene technology has further improved manufacturing processes and enabled the commercialization of enzymes that could previously not be produced on large scale. Furthermore, modern biotechnologies, such as protein engineering and directed evolution, have further revolutionized the development of industrial enzymes (see Section 7.3). These advances have made it possible to provide tailor-made enzymes displaying new activities and adapted to new process conditions, enabling a further expansion of their industrial use.

Table 1 (adapted from F. Hasan et al.) illustrates the estimated value of the worldwide use of industrial enzymes and the market segmentation. The technical industries,

dominated by the detergent, starch, textile and fuel alcohol industries, account for the major consumption of industrial enzymes.

	2002	2003	2004	2009
Technical enzymes	978.2	1009.2	1040.0	1222.0
Food enzyme	701.0	720.0	740.0	863.0
Animal feed enzyme	210.8	215.6	220.0	267.0
Total	1890.0	1945.0	2000.0	2352.0

Table 1. Global enzyme markets by application sectors, through 2009 (\$ millions). Adapted from F. Hasan et al. / *Enzyme and Microbial Technology* 39 (2006) 235–251

Table 2 reports the more important applications of industrial enzymes. Most of the industrial enzymes are hydrolases and are used for the degradation of various natural substances (<http://www.amano-enzyme.co.jp/english/index.html>). Proteases are traditionally among the most widely employed enzymes, because of their extensive use in the detergent and dairy industries. Glycosidases (amylases and cellulases) are largely used in the textile, detergent and baking industries; in the last few years the production of amylases and cellulases has dramatically improved because of their use in the bioethanol industries as catalysts for the saccharification of starch and cellulose.

Industry	Enzyme class	Application
Detergent (laundry and dish wash)	Protease	Protein stain removal
	Amylase	Starch stain removal
	Lipases	Lipid stain removal
	Cellulase	Cleaning, color clarification, anti-redeposition (cotton)
	Mannanase	Mannanan stain removal (reappearing stains)
Starch	Amylase	Starch liquefaction and saccharification
	Cellulase	Cellulose saccharification
	Xylanase	Viscosity reduction
	Pullulanase	Saccharification
	Glucosidase	Saccharification
	Glucose isomerase	Glucose to fructose conversion
	Cyclodextrin-glycosyltransferase	Cyclodextrin production
Food (including dairy)	Protease	Milk clotting, infant formulas (low allergenic), flavor
	Lipase	Cheese flavor

	Lactase (β -galactosidase)	Lactose removal (milk)
	Pectin methyl esterase	Firming fruit-based products
	Pectinase	Fruit-based products
	Transglutaminase	Modify visco-elastic properties
Baking	Amylase	Bread softness and volume, flour adjustment
	Xylanase	Dough conditioning
	Lipase and phospholipase	Dough stability and conditioning (<i>in situ</i> emulsifier)
	Glucose oxidase	Dough strengthening
	Lipoxygenase	Dough strengthening, bread whitening
	Protease	Biscuits, cookies
	Transglutaminase	Laminated dough strengths
Beverage	Pectinase	De-pectinization, mashing
	Amylase	Juice treatment, low calorie beer
	β -Glucanase	Mashing
	Acetolactate decarboxylase	Maturation (beer)
	Laccase	Clarification (juice), flavor (beer), cork stopper treatment
Textile	Cellulase	Denim finishing, cotton softening
	Amylase	De-sizing
	Pectate lyase	Scouring
	Catalase	Bleach termination
	Laccase	Bleaching
	Peroxidase	Excess dye removal
Pulp and paper	Lipase	Pitch control, contaminant control
	Protease	Biofilm removal
	Amylase	Starch-coating, de-inking, drainage improvement
	Xylanase	Bleach boosting
	Cellulase	De-inking, drainage improvement, fiber modification
Fats and oils	Lipase	Transesterification
	Phospholipase	De-gumming, lyso-lecithin production
Leather	Protease	Unhearing, bating

Personal care	Lipase	De-pickling
	Amyloglucosidase	Antimicrobial (combined with glucose oxidase)
	Glucose oxidase	Bleaching, antimicrobial
	Peroxidase	Antimicrobial

Table 2. Examples of applications of enzymes in industries different from chemical and pharmaceutical (adapted from <http://www.novozymes.com>)

3. Advantages and Drawbacks Related to the Use of Biocatalysts in Chemistry

Many enzymes have been found to catalyze a variety of reactions that can be very different from the reaction and substrate with which the enzyme is associated in nature.

In many cases biocatalysts respond to the needs of technological solutions coming from the transformation industry, as many classical chemical transformation processes have inherent drawbacks from a commercial and environmental point of view. As a matter of fact, non-specific reactions may result in poor product yields. High temperatures and/or pressures needed to drive reactions leads to high energy costs and may require large volumes of cooling water downstream. Harsh and hazardous processes involving high temperatures, pressures acidity or alkalinity need high capital investment and specially designed equipment and control systems. Unwanted by-products may prove difficult or costly to dispose of. High chemical and energy consumption as well as harmful by-products have a negative impact on the environment.

Compared with traditional methods, biocatalysis often offers a number of advantages such as:

- High stereo-, regio- and chemoselectivity
- Decreased requirements of tedious protection and de-protection schemes
- Lower incidence of by-products
- Mild reaction conditions
- Efficient catalysis of both simple and complex transformations
- Uncomplicated and cheap refining and purification
- Reduced impact of manufacturing on the environment by reducing the consumption of chemicals and energy, and the subsequent production of waste.

Finally, it must be underlined that only small amounts of enzymes are required to carry out chemical reactions even on an industrial scale.

Most of these advantages are potential; for example stereoselectivity is not often guaranteed, especially when synthetic substrates are employed.

In order to judge the environmental impact of a process, the *E* factor can be used (Margreth et al., 2001), which is the ratio between the mass of waste material and the mass of desired products.

$$E\text{-Factor} = \frac{\text{Mass of waste material}}{\text{Mass of products}}$$

The *E* factor allows to compare processes and designate the amount of by-products (e.g. solvent losses, acids and bases used in work-up, process aids, waste from energy production), produced per kg of product. In this context, waste will include any reaction product that does not have any further use, and also reagents and solvents used during the course of manufacture that are not re-used or recycled. The *E*-factors vary enormously between the different sectors of the chemical industry as reported in Table 2.4: It increases substantially from bulk to fine chemicals and specialties. This is partly due to the fact that the production of fine chemicals generally involves multi-step syntheses and partly to the widespread use of stoichiometric rather than catalytic reagents, as in the case of biocatalysis.

Industry	Product tonnage	Kg by-product / kg product
Oil refining	10 ⁶ - 10 ⁸	<<0.1
Bulk chemical	10 ⁴ - 10 ⁶	< 1-5
Fine chemical	10 ² - 10 ⁴	5-50
Pharmaceuticals	10- 10 ³	25->100

Table 3. Relation between the *E* factor and sectors of industry (Adapted from Wegman et al.).

A “good” *E*-factor would typically be around 0.1 so that 10 kg of desired product produces 1 kg of waste and by-product. At the other extreme, in pharmaceutical manufacturing when a high-purity is essential, the *E*-factor can be as high as 100, meaning that 1 kg of product produces 100 kg of waste. These figures indicate that new, more efficient synthetic methodologies for the production of pharmaceuticals are urgently required. As an example, the translation of the productive processes of β -lactam antibiotics to biocatalysis allowed a 5-fold reduction of the corresponding *E*-factor (see Section 12.2).

Besides the above mentioned advantages, some drawbacks coming from the application of biocatalysis in chemistry must be recognized, such as:

- Biocatalysts often show lower stability than conventional catalyst
- Development of industrial biocatalytic processes are usually much longer to establish
- Low number of commercially available biocatalysts
- Necessity of microbiological facilities if the biocatalyst is not a commercial enzyme

Most of these disadvantages might be overcome by modern techniques (screening, molecular biology, protein engineering, immobilization) able to furnish a much higher number of biocatalysts with improved performances.

-
-
-

TO ACCESS ALL THE 40 PAGES OF THIS CHAPTER,
Visit: <http://www.eolss.net/Eolss-sampleAllChapter.aspx>

Bibliography

Aehle W. (2004) *Enzymes in Industry 2nd Edition*, Wiley VCH, Weinheim, Germany. [A broad overview of the use of enzymes in all the industrial fields]

Arnold F. H. (2001) Combinatorial and computational challenges for biocatalyst design. *Nature* **409**, 253-257. [A review illustrating the potential of laboratory evolution methods for tuning enzyme properties]

Bommarius A.S. and Riebel B. (2004) *Biocatalysis* Wiley-VCH, Weinheim, Germany. [A milestone book covering all the aspects of biocatalysis, from fundamentals to applications]

Bouzas T.D., Barros-Velazquez J., Villa T.G. (2006) Industrial applications of hyperthermophilic enzymes: A review. *Protein and peptide letters* **13**, 445-451. [A review about the discovery and application of enzymes produced by thermophilic and hyperthermophilic microorganisms]

Braiuca P., Ebert C., Basso A., Linda P., Gardossi L. (2006) Computational methods to rationalize experimental strategies in biocatalysis. *Trends in Biotechnology* **24**, 419-425. [A critical review on how computational methods can be successfully applied to biocatalysis]

Cantone S., Hanefeld U., Basso A. (2007) Biocatalysis in non-conventional media-ionic liquids, supercritical fluids and the gas phase *Green Chemistry* **9**, 954-971 [A survey of recent developments of biocatalysis in non-conventional media other than organic solvents]

Cao L. (2006) *Carrier-bound Immobilized Enzymes: Principles, Application and Design* Wiley-VCH, Weinheim, Germany. [A systematic overview of the techniques of immobilization of enzymes including all recent developments]

Davies G., Henrissat, B. (1995). Structures and mechanisms of glycosyl hydrolases. *Structure* **3**, 853-859. [A comprehensive discussion about the mechanisms and the stereochemistry of glycosyl hydrolases]

Dean S.M., Greenberg W.A., Wong C.H. (2007) Recent advances in aldolase-catalyzed asymmetric synthesis *Advanced Synthesis and Catalysis* **349**, 1308-1320. [An overview on the recent advances in stereoselective biotransformations with aldolases]

Gadler P., Glueck S.M., Kroutil, W., Nestl, B.M., Larslegger-Schnell B., Uerbacher B.T., Wallner S.R., Faber K. (2006) Biocatalytic approaches for the quantitative production of single stereoisomers from racemates. *Biochemical Society Transactions* **34**, 296-300. [A discussion about how to transform a racemate into a single stereoisomeric product in quantitative yield]

Halling P.J. (2000) Biocatalysis in low-water media: understanding effects of reaction conditions. *Current Opinion in Chemical Biology* **4**, 74-80. [A critical review on the advances in the field of non-aqueous biocatalysis]

Halling P.J., Ulijn R.V., Flitsch S.L. (2005) Understanding enzyme action on immobilised substrates *Current Opinion in Biotechnology* **16**, 385-392. [An exhaustive review on the fundamental theoretical as well as on some applicative issues related to this technology]

Hasan F., Shah A.A. and Hameed A. Industrial applications of microbial lipases. (2006) *Enzyme and Microbial Technology* **39**, 235–251. [A review concerning the industrial use of one the most used class of enzymes]

Klibanov A.M. (2001) Improving enzymes by using them in organic solvents. *Nature* **409**, 241-246.

Koeller K.M. and Wong C.H. (2001) Enzymes for chemical synthesis. *Nature* **409**, 232-241. [A general survey of the features of enzymatic methods applied for solving chemical problems]

Leisola M., Turunen O. Protein engineering: opportunities and challenges. *Applied Microbiology and Biotechnology* **75**, 1225-1232. [An overview of the rapid progress and impressive results in the field of protein engineering using rational design and random techniques or a combination of both]

Liese A., Seelbach K., Wandrey C. 2006. *Industrial Biotransformations 2nd Edition* Wiley-VCH, Weinheim, Germany. [A systematic description of biotransformations of industrial interest]

Lombard C., Saulnier J., Wallach J.M. (2005) Recent trends in protease-catalyzed peptide synthesis. *Protein and Peptide Letters* **12**, 621-629. [A clear dissertation on thermodynamically- and kinetically-controlled enzymatic synthesis of peptides]

Molinari F. (2006) Oxidations with isolated and cell-bound dehydrogenases and oxidases. *Current Organic Chemistry* **10**, 1247-1263. [An overview of the oxidative potential of oxidoreductases]

Nakamura K, and Matsuda T. (2006) Biocatalytic reduction of carbonyl groups. *Current Organic Chemistry* **10**, 1217-1246. [The potential and the applications of enzymatic reduction of prochiral carbonyls]

Patel R.N. (2006) Synthesis of chiral intermediates for pharmaceuticals. *Current Organic Chemistry* **10**, 1289-1321. [A description of biocatalytic processes for the synthesis of optically pure chiral intermediates for the synthesis of pharmaceuticals]

Pollard D.J., Woodley J.M. (2007) Biocatalysis for pharmaceutical intermediates: the future is now. *Trends in Biotechnology* **25**, 66-73. [An updated overview on the impact of biocatalysis in the pharmaceutical industry]

Trost B.M. (2004) Asymmetric catalysis: an enabling science. *Proceedings of the National Academy of Sciences USA* **101**, 5348-5355. [On the importance of developing new efficient methods for producing optically pure compounds]

Ulijn R. V., De Martin L., Gardossi L., Halling P.J., Biocatalysis in reaction mixtures with undissolved solid substrates and products, *Current Organic Chemistry* **2003**, *7*, 1333-1346. [A review describing the theoretical basis as well the application of biocatalysis on undissolved substrates]

Urlacher V.B., Lutz-Wahl S., Schmid R.D. (2004) Microbial P450 enzymes in biotechnology. *Applied Microbiology and Biotechnology* **64**, 317-325. [The applications in biocatalysis of this extraordinary class of oxygenases]

Wegman M.A., Janssen M.H.A., van Rantwijk F., Sheldon R.A. (2001). Towards Biocatalytic synthesis of β -lactam antibiotics. *Advanced Synthesis and Catalysis* **343**, 559-576. [A case-study concerning the replacement of traditional chemical procedures with biotransformations, driven by the desire to reduce waste and the dependence on organic solvents]

Wijffels R.H. (2001) *Immobilized Cells* Springer-Verlag, Heidelberg, Germany. [A handbook describing the techniques of immobilization of whole cells including discussions and recent applications]

Relevant web-sites

<http://www.brenda-enzymes.info> [A comprehensive enzyme information system with structural and functional data]

<http://www.novozymes.com> [The web-site of the major enzyme producer (Novozymes, Denmark), providing information on enzyme industrial applications]

<http://www.amano-enzyme.co.jp/english/index.html> [The web site of the major Japanese enzyme producer, with commercial and technical information on biocatalysts]

Biographical Sketches

Lucia Gardossi was born in Trieste, Italy in 1963 and graduated in Pharmaceutical Chemistry and Technology in 1988 at the University of Trieste. Since the beginning of her scientific activity she has always investigated the application of computational chemistry to the understanding and development of novel biocatalyzed systems. She got the PhD in Medicinal Chemistry at the University of Trieste working on enzymatic modification of peptides. From 1989 to 1991 she worked at the Massachusetts Institute of Technology (Cambridge, USA) in the group of Prof. A.M. Klibanov studying the use of enzymes in non-conventional media. From 1993 to 1995 she studied enzymatic modification of carbohydrates at POLY-TECH Soc. coop. (Trieste- Italy).

Currently she is associate professor of organic chemistry at the Faculty of Pharmacy of the University of Trieste. She is author of more than 50 publications in the field of biocatalysis and 3 patents. She is Italian delegate inside the European Section of Applied Biocatalysis (ESAB) of the European Federation of Biotechnology (EFB). She is member of the scientific board of the Italian Technology Platform on Sustainable Chemistry and of the Italian Association of Biocatalysis and Bioseparations (AIBB).

Her current research activity is focused on different aspects of biocatalysis: enzymes in non-conventional media, computational methods for the prediction of enzyme selectivity/stability, immobilization of enzymes, chemo-enzymatic synthesis on solid phase, development of novel supports for solid phase synthesis and for enzyme immobilization, modeling of thermodynamics of bio catalyzed reactions.

Francesco Molinari was born in 1961 in Milano, Italy. He studied chemistry and graduated from the University of Milano, Italy with a Ph.D. thesis about "Supramolecular Chemistry: some applications to covalent bond cleavage and formation" in 1992 under the supervision of Prof. Cesare Gennari. During these years his fields of interest have been organic synthesis applied to the problems of molecular recognition, bifunctional catalysis and stereoselective carbon-carbon formation. He was a postdoctoral fellow at the Instituto Tecnico of Lisbon working on extractive bioconversions. He began his independent career at the Industrial Microbiology Section of the Department of Food Science and Microbiology, University of Milano in 1992. Since 2000 he has been Professor of Biotechnology and Chemistry of Fermentations at the University of Milano. Professor Molinari's research interests include (stereo)-selective biotransformations, production and isolation of microbial secondary metabolites and wine fermentations.

He has been member of the Scientific Board of the Italian Society of General Microbiology and Microbial Biotechnology (SIMGBM) and of the Italian Association of Biocatalysis and Bioseparations (AIBB).