

SEASONAL AND SOCIAL FACTORS AFFECTING REPRODUCTION

Rodolfo Ungerfeld

Departamento de Fisiología, Facultad de Veterinaria, Universidad de la República, Montevideo, Uruguay

Alejandro Bielli

Departamento de Morfología y Desarrollo, Facultad de Veterinaria, Universidad de la República, Montevideo, Uruguay

Keywords: anestrus, biostimulation, buck effect, male effect, estrous cycle, ovulation, pheromones, photoperiod, ram effect, reproductive patterns, ruminants, seasonal breeding, social cues, social influences on reproduction, swine

Contents

1. Introduction
2. Environmental regulation of reproduction in mammals
 - 2.2 Seasonal reproductive strategies
 - 2.3. Extrinsic and intrinsic factors and ultimate and proximate factors influencing seasonal reproduction
 - 2.3.1. Ultimate factors
 - 2.3.2. Energy balance
 - 2.3.3. Nutrients
 - 2.3.4. Competence for food and predation pressure
 - 2.3.2. Proximate factors:
- 2.4. Mechanisms regulating seasonal reproduction with photoperiod as a predictor
- 2.5. Seasonal reproduction in livestock species
 - 2.5.1. Sheep
 - 2.5.2. Goats
 - 2.5.3. Cattle
 - 2.5.4. Swine
3. Socio-sexual signals
 - 3.1. The ram effect
 - 3.1.1. Managing the ram effect
 - 3.1.2. The ram effect in cyclic ewes
4. Socio-sexual signals and seasonal reproductive patterns
5. General overview

Summary

The present chapter summarizes the current information on how some environmental factors, mainly seasonality and socio-sexual signals, interact with the endogenous mechanisms that control reproductive activity in domestic ruminants and swine. Ultimate factors such as latitude and seasonality, energy balance, nutrients, competence for food and predation pressure, as well as proximate factors such as nutrition, temperature, and socio-sexual influences are explained in relation to the main predictor:

photoperiod. The main seasonal patterns of the domestic species included in the chapter are presented. This review also includes an update on socio-sexual signals, with sheep as reference model (the ram effect), and how these signals can be included in field practices to induce out-of-season births. Although still scarce, information on the use of the ram effect in cyclic ewes is updated. Lastly, the information on how socio-sexual signals can alter seasonal reproductive patterns is presented.

1. Introduction

The reproductive patterns of an animal are a consequence of the interaction of the endogenous regulatory mechanisms – mainly endocrine– and environmental signals. These signals may affect reproductive mechanisms deeply, i.e., females can stop cycling in some periods of the year, or due to the presence of a dominant individual on the group, or can ovulate and come into estrus during the seasonal non-cycling period due to the acute stimulus of male presence.

2. Environmental Regulation of Reproduction in Mammals

It is well known that the expression of a given trait (phenotype) in an animal depends on the combined influence of both, genetic factors (genotype) and environmental factors. The main environmental factors influencing animal reproduction are temperature, humidity, amount and distribution of rainfall, solar radiation and photoperiod, nutrition, productive system management, social interactions among individuals within the same population, predator-prey interactions, parasite- and pathogen- host interactions (Sadleir, 1968; Giwercman and Giwercman, 2011, Taberlet et al., 2011, Burns et al., 2010).

Most wild mammals, especially those of greater size and longevity, are to some extent, seasonal breeders (Goldman et al., 2004). Such animals limit their mating activity and offspring births to well defined seasons of the year. However, some domestic species such as cattle, pigs and rabbits exhibit no seasonal breeding if they are raised in environments with mild climatic changes throughout the year.

2.2 Seasonal Reproductive Strategies

Reproduction is a metabolically highly demanding process, and generally offspring are quite more sensitive to deleterious environmental factors than their parents (Foster and Nagatani, 1999). Reproductive strategies vary with the genetic background of different animal species, and the most important environmental factors are those having greater influence on offspring survival. Many mammalian wild species inhabiting temperate zones adjust their reproductive season so that offspring births are concentrated during spring. Available resources are most scarce in winter, and offspring being born in spring have better chances of survival the older and heavier they are when they will be forced to face wintertime conditions. On the other hand, it seems to be easier, from an evolutionary perspective, to change the timing of the mating season than to change the duration of gestation or lactation.

The degree of reproductive seasonality expressed by any animal can vary markedly in intensity and timing. Variations are driven according to the above mentioned environmental factors, the species to which it belongs, its gender, its particular genotype. However, variations are also driven accordingly to many factors pertaining to the individuals' life history, especially their reproductive status, and amount of body energetic stores.

2.3. Extrinsic and Intrinsic Factors and Ultimate and Proximate Factors Influencing Seasonal Reproduction

Factors having an influence on seasonal reproduction in any animal can be classified as intrinsic and extrinsic factors (Bronson, 1989). While intrinsic factors are related to genotype, and belong to the individual itself, extrinsic factors are related to the animal's environment. An example of intrinsic factor is life expectancy, as related to annual environmental variations: in an animal having a life expectancy of approximately one year or less, seasonality will be barely expressed, and the animal will develop opportunistic breeding strategies. Alternatively, an animal having a life expectancy of several years will have more chances of giving birth to viable offspring when restricting births to the most favorable seasons of the year. Similarly, the bigger the body size, the higher the chances of expressing seasonal reproduction, since a bigger animal has bigger energetic body stores, and can postpone easily its breeding activity to seasons more favorable for the newborn. Body size is, however, less important than both life expectancy and diet flexibility: an animal depending on a reduced spectrum of food will generally breed much more seasonally than an animal of similar traits but which is able to feed on a more flexible and varied diet. Another relevant intrinsic factor is gender. Generally speaking, males will tend to have longer breeding seasons than their female counterparts, since spermatogenesis commonly requires significantly more time to complete than ovarian follicle maturation and ovulation (Simpson et al., 1982, Schlatt et al., 1995), and masculine earlier recrudescence of breeding activity may facilitate intrasexual competition for mates (Prendergast, 2005).

Environmental (extrinsic) factors which are decisive on reproductive activity can be classified as ultimate and proximate factors, according to the time when they act on breeding activity. Ultimate factors are important in the long term, from an evolutionary point of view. Generally, the most important ultimate factor is food availability and its influence on energetic balance (Bronson, 2009), as reproduction is energetically highly demanding. The costlier period in mammals extends from late gestation to the first part of lactation. These breeding activities will suffer a strong selective pressure to occur in seasons of the year when most food is available, generally in spring and summer. Other ultimate factors which might be of importance are intra- and inter-species competition for resources, as well as strategies aiming at avoiding or limiting predation on offspring.

2.3.1. Ultimate Factors

2.3.1.1. Latitude and Seasonality

High latitudes display great annual variations in photoperiod and ambient temperature. Most animals adapted to such climates have short, well-defined breeding seasons,

whereas tropical latitudes, where photoperiod and temperature vary little along the year, are inhabited by many more animals with long breeding seasons, and many tropical animals breed all year round (Bronson, 1988). Thus, for any given species, the breeding season generally prolongs as latitude diminishes. The latitude at which reproduction becomes annual diminishes as the animal's life expectancy increases. Furthermore, climatic conditions do vary annually. Animals with short life expectancy vary more or less markedly their seasonal reproductive cycles annually, according to prevalent conditions. Conversely, inter-annual variations in reproductive activity tend to be smaller in animals with long life expectancy.

2.3.2. Energy Balance

Generally speaking, the periods of the year when food availability becomes a bottleneck are very important as driving forces for seasonal reproduction. This is the ultimate cause of seasonal reproduction in all mammals and the proximate cause in many other species (review: Bronson, 2009). Interestingly, the smaller the size of the mammal, the more important tends to be the lactational bottleneck, whereas as body size increases, post weaning food availability generally becomes the main bottleneck. In most ancestors of livestock species, weaning food availability was very important.

2.3.3. Nutrients

Besides energetic balance, availability of different key nutrients varies throughout the year. This fact can become, for some species, an important limiting factor to seasonal reproduction.

2.3.4. Competence for Food and Predation Pressure

Several species which are competing totally or partially for the same resource (i.e. pasture) can be strongly selected for desynchronization of their breeding seasons. Such a situation has been well documented for herbivorous ungulates in the African savanna (Mossman & Mossman, 1962).

2.3.2. Proximate Factors:

Proximate factors are responsible for immediate influences on breeding activity.

2.3.2.1. Nutrition

Nutrition is a very important factor. It can act not only as a paramount ultimate factor, but as a proximate factor as well in many cases. Furthermore, if food availability falls below a threshold that may vary according to species and populations, an animal will not breed. Moreover, changes in food availability can influence seasonal breeding patterns. This effect has been extensively studied in sheep, and marked differences in sensitivity to food availability have been described. Generally speaking, the effect of food availability on seasonal breeding is much stronger in breeds originated from Mediterranean or tropical climates than in breeds originated from temperate latitudes (Martin and Walkden-Brown, 1995, Boukhliq et al., 1996, Bielli et al, 1999, Zarazaga et

al, 2005). Conversely, food availability or even some increase in nutrient availability can be the proximate factor triggering breeding activity.

2.3.2.2. Temperature

Ambient temperature can also be a proximate factor, since warmer temperatures could trigger reproductive activity. Conversely, too high temperatures could limit reproductive activity (Haim et al., 2005).

2.3.2.3. Social Factors

The reproductive activity of fellow individuals can trigger reproductive activity of others within the same population. This is going to be dealt with in the second part of the present chapter.

2.3.2.4. Predictors: Photoperiod

The most widespread proximate factor timing seasonal reproduction is the annual photoperiodic variation (Ortavant et al., 1988). It is well known that due to the inclination of the Earth's rotation axis and Earth's translation along its orbit, the duration of daylight (photoperiod) varies along the year, and that such variation is more marked nearer the poles than near the Equator. Photoperiod is a very reliable predictor of future environmental conditions, since it is a very constant environmental clue. Furthermore, it predicts seasonal changes in climate and food availability (Bradshaw and Holzapfel, 2007). Some species (i.e., sheep, goat, deer) are known as 'short-day breeders' because their breeding season occurs mainly when days are getting shorter (summer and autumn). Such species normally have gestation lengths of 5-6 months. Thus, their offspring are usually born in spring. On the other hand, other species having either short gestation lengths (1-2 months, i.e., hamster, mink) or long gestation lengths (horse) are 'long-day breeders', and breed in late winter or spring, with their offspring also being born mainly in spring.

2.4. Mechanisms Regulating Seasonal Reproduction with Photoperiod as a Predictor

Most seasonal breeding mammals studied have been shown to possess a self-sustained endogenous rhythm of seasonal reproductive activity (circannual biological clock, reviews: Rensing et al., 2001; Lincoln et al., 2006), which is either synchronized or entrained by photoperiod (review: Kumar, 1997). Daylight is perceived essentially by the eyes. The retina of the eye is stimulated by light and transmits neural information through the optic nerve to the suprachiasmatic nucleus (a group of neurons located in the ventral side of the encephalon). The information is transmitted to the stellate ganglion (anterior cervical ganglion) and then to the pineal gland (Bittman et al., 1983). The pineal gland is a small endocrine gland in the brain. Its shape is similar to a pine cone (hence its name). The pineal gland apparently evolved from a photoreceptor organ which used to be situated on the roof of the brain in primitive vertebrates (Ekström and Meissl, 2003). It is pivotal in mediating the effects of photoperiod on reproduction, as it

translates neural stimuli mediating photoperiod into hormonal stimuli. The hormone secreted by the pineal gland is melatonin (N-acetyl-5-methoxytryptamine).

Melatonin is a small indole compound (chemically related to aminoacids) secreted by the pineal gland when the animal is in a dark environment. Melatonin secretion is inhibited by daylight. High melatonin concentrations in blood transmit the information that the animal is in a dark environment to the animal's organs and tissues. Thus, melatonin can either inhibit ('long day breeders') or stimulate ('short day breeders') reproductive activity (Figure 1).

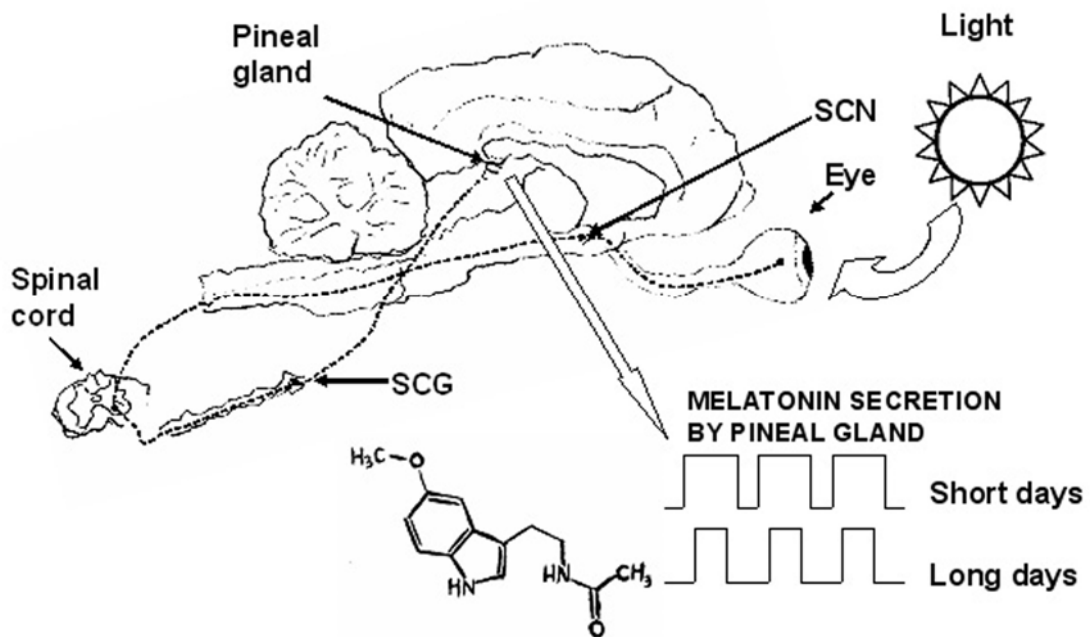


Figure 1. Regulation of pineal gland melatonin secretion by photoperiod: schematic sagittal view of a sheep's eye and brain. Light is perceived by the eye's retina (i.e., photic information is converted into neural information). Neural information generated in the retina is transmitted to the suprachiasmatic nuclei (SCN). Connections from the SCN reach the spinal cord. Spinal neurons send the information to the anterior cervical ganglion (ACG). Postganglionic SCG neurons send neural paths to the pineal gland, where neural information is converted into hormonal information (high melatonin secretion levels during dark hours). The duration of high melatonin blood levels indicates the duration of dark hours to the whole animal's body. Melatonin is a small indole molecule (bottom of figure).

2.5. Seasonal Reproduction in Livestock Species

2.5.1. Sheep

Most sheep breeds show reproductive seasonality, normally breeding in late summer and autumn. Because of genetic differences, different breeds of sheep maintain different levels of photo-responsiveness (Gómez-Brunet et al., 2008). This is one reason why differences of breeding time occur between different breeds of sheep. Seasonal

reproduction occurs widely in mid and high-latitude breeds, but is very weak or does not exist at all in breeds originated near the Equator, i.e., most hair sheep breeds.

Both rams and ewes have seasonal variations in breeding activity, but females are more seasonal than males, and ewes' breeding season is normally shorter than rams' one, from the same breed. Temperature, nutrition, social influences, lambing date and lactation period can modulate photo-periodical influences (Scaramuzzi and Martin, 2008; Forcada and Abecia, 2006). It is possible to manipulate breeding season timing by altering the photoperiod with artificial lightning or with melatonin implants that simulate melatonin blood concentrations similar to the breeding season.

2.5.2. Goats

Goats are also well known as seasonal breeders. Seasonal patterns are similar to those of sheep. The beginning and duration of the breeding season is dependent on several factors such as latitude, climate, breed, physiological stage, presence of the male (buck effect), breeding system and photoperiod. In temperate regions, goats breeding season occurs in autumn and winter. In tropical regions, goats are normally continuous breeders, but anestrus periods are frequent when food is scarce. Generally speaking, tropical origin breeds are less seasonal or not seasonal at all. Interestingly, melatonin implants are markedly more effective in goats than in sheep breeds of Northern Europe (Chemineau et al., 1992). Photoperiodic treatments coupled with buck effect allow synchronization of ovulation but fertility results are poorer than those of hormonal treatments (Fatet et al., 2011).

2.5.3. Cattle

Cattle breeds originated from temperate climates are normally not seasonal breeders. However, ancestral species of domestic cattle are seasonal breeders, and cows living in very high latitudes do breed seasonally, with calving concentrating in spring (Borisenkov et al., 2004). Furthermore, there are mild seasonal variations in hormone secretion in bulls (Stumpf et al., 1993). Thus, it seems that domestication and selection for all year round breeding has been successful in cattle, but still a hidden endogenous rhythm exists, which can manifest itself under extreme conditions.

2.5.4. Swine

Female pig ancestors (wild European boar) are seasonal polyestrous breeders, with two reproductive periods: the main period extends from November to March in the Northern Hemisphere, and the secondary from April to May (Mauget, 1972; Mauget et al., 1984). Feral pigs adapt easily to seasonal breeding. However, seasonal breeding patterns of domestic pigs are more flexible with the duration and timing of their breeding seasons (Ravault et al., 1982). In the domestic pig seasonal variations in prolificacy still exist. Boars not only show decreased steroid synthesis, sperm counts and libido in summer when compared with the winter but also show a biphasic pattern with a transient increase in spring. In cyclic sows anestrus may appear mainly in summer and occasionally in February/March (Northern Hemisphere) (Claus and Weiler, 1985). Periods of infertility and late pregnancy loss are frequent in both sows and gilts during

summer (Bertoldo et al., 2009), and periods of early pregnancy disruption exist along summer and early autumn (Tast et al., 2002). However, swine can normally breed throughout the year if food and housing are adequate (Macchi et al., 2010). Both hormonal melatonin treatments and photoperiod manipulation are effective, but not a sustainable solution to seasonal infertility, mainly because of economic reasons (Bassett et al., 2001).

-
-
-

TO ACCESS ALL THE 24 PAGES OF THIS CHAPTER,
Visit: <http://www.eolss.net/Eolss-sampleAllChapter.aspx>

Bibliography

Bassett J.M., Bray C.J., Sharpe C.E. (2001). Reproductive seasonality in domestic sows kept outdoors without boars. *Reproduction* 121, 613–629. [Interesting paper on the effects of melatonin implants and the decrease of sows' seasonal anestrus].

Bertoldo M., Grupen C.G., Thomson P.C., Evans G., Holyoake P.K. (2009). Identification of sow-specific risk factors for late pregnancy loss during the seasonal infertility period in pigs. *Theriogenology* 72, 393–400. [Report on late pregnancy loss as a manifestation of seasonal infertility in pigs, and the risk factors that producers can manipulate during the seasonal infertility period to improve breeding herd productivity].

Bielli A., Pedrana G., Gastel M.T., Castrillejo A., Moraña A., Lundeheim N., Forsberg M., Rodriguez-Martinez H. (1999). Influence of grazing management on the seasonal change in testicular morphology in Corriedale rams. *Animal Reproduction Science* 56, 93–105. [Report on nutritional influences, grazing management and the partial decrease of seasonal testis growth cycle in Corriedale rams].

Bittman E.L., Karsch F.J., Hopkins J.W. (1983). Role of the pineal gland in ovine photoperiodism: regulation of seasonal breeding and negative feedback effects of estradiol upon luteinizing hormone secretion. *Endocrinology* 113, 329–336. [This is a pioneer report which determined that the pineal gland mediates photoperiodic control of seasonal breeding in ewes].

Borisenkov M.F., Kalinin A.I., Vakhnina, N.A. (2004). Seasonal dynamics of cattle reproduction in the North. *Russian Journal of Ecology* 35, 205–208. [Short paper studying the circannual rhythm of cow reproduction in Northern regions]

Boukhliq R., Adams N.R., Martin G.B. (1996). Effect of nutrition on the balance of production of ovarian and pituitary hormones in ewes. *Animal Reproduction Science* 45, 59–70. [A relevant report of nutritional influences on the balance between pituitary FSH secretion and gonadal feedback by changing the responsiveness to the inhibitory effects of estradiol and inhibin].

Bradshaw W.E., Holzapfel C.M. (2007). Evolution of animal photoperiodism. *Annual Reviews of Ecology and Evolution Systems* 38, 1–25. [A comprehensive review of the ability of organisms to assess and use the day length as an anticipatory cue to time seasonal events in their life histories. It deals with a wide range of organisms, from rotifers to mammals].

Bronson F.H. (1988). Mammalian reproductive strategies: genes, photoperiod and latitude. *Reproduction Nutrition Development* 28, 335–347. [An interesting review which considers how and why natural selection might promote or block the photoperiodic regulation of a mammal's reproduction].

Bronson F.H. (1989). Mammalian reproductive biology. Chicago, IL: The University of Chicago Press. pp. 51–52. [A very interesting, easy-to-read book. It is a comprehensive, interdisciplinary overview of the way mammals reproduce and the strategies they have evolved in order to achieve reproductive success].

Bronson F.H. (2009). Climate change and seasonal reproduction in mammals. *Philosophical Transactions of the Royal Society of London B Biological Sciences* 364, 3331–3340. [This paper (i) discusses the neuroendocrine pathways via which foraging conditions and predictive cues such as photoperiod enforce seasonality, (ii) considers the kinds of seasonal challenges mammals actually face in natural habitats, and (iii) uses the information thus generated to suggest how seasonal reproduction might be influenced by global climate change.].

Bruce H.M. (1959). An exteroceptive block to pregnancy in the mouse. *Nature* 184, 105. [A pioneer paper on the existence of pheromones in mammals]

Burns B.M., Fordyce G., Holroyd R.G. (2010). A review of factors that impact on the capacity of beef cattle females to conceive, maintain a pregnancy and wean a calf-Implications for reproductive efficiency in northern Australia. *Animal Reproduction Science* 122, 1–22. [This is a review of factors that may impact on the capacity of beef cattle females, grazing semi-extensive to extensive pastures in northern Australia, to conceive, maintain a pregnancy and wean a calf. Conclusions and recommendations to minimize reproductive inefficiencies based on current knowledge are presented]

Chemineau P. (1983). Effect on oestrus and ovulation of exposing creole goats to the male at three times of the year. *Journal of Reproduction and Fertility* 67, 65–72. [An initial report on the effect of buck introduction in goats, comparing possible effects according to the year period]

Chemineau P., Gauthier D., Poirier J.C., Saumande J. (1982). Plasma levels of LH, FSH, prolactin, oestradiol-17beta and progesterone during natural and induced oestrus in the dairy goat. *Theriogenology* 17, 313–323. [An article characterizing the endocrine and ovarian responses to the buck effect]

Chesworth J.M., Tait A. (1974). A note on the effect of the presence of rams upon the amount of luteinizing hormone in the blood of ewes. *Animal Production* 19, 107–110 [One of the earlier studies determining that the presence of rams stimulates LH secretion in the ewe].

Claus R., Weiler U. (1985). Influence of light and photoperiodicity on pig prolificacy. *Journal of Reproduction and Fertility, Suppl.* 33, 185–197. [This is a review of environmental factors that influence on the capacity of pigs to overcome seasonal restraints to pig reproductive success].

Cognie Y., Gray S.J., Lindsay D.R., Oldham C.M., Pearce D.T., Signoret J.P. (1982). A new approach to controlled breeding in sheep using the “ram effect”. *Proceedings of the Australian Society of Animal Production* 14, 519–522. [This article presents some experiments on the use of the ram effect, including the use of progestagen primings]

Contreras-Solis I., Vasquez B., Diaz T., Letelier C., Lopez-Sebastian A., Gonzalez-Bulnes A. (2009). Efficiency of estrous synchronization in tropical sheep by combining short-interval cloprostenol-based protocols and “male effect”. *Theriogenology* 71, 1018–1025. [An article combining the use of the ram effect with traditional estrus synchronization treatments]

Cushwa T.W., Bradford G.E., Stabenfeldt G.H., Berger Y.M., Dally, M.R. (1992). Ram influence on ovarian and sexual activity in anestrus ewes: effects of isolation of ewes from rams before joining and date of ram introduction. *Journal of Animal Science* 70, 1195–1200. [A study that aimed to determine the importance of ewes’ isolation before the ram effect]

Delgadillo J.A., Gelez H., Ungerfeld R., Hawken P.A.R., Martin G.B. (2009). Revisiting the dogmas surrounding the mechanisms involved in the male effect in sheep and goats. *Behavioural Brain Research* 200, 304–314. [A review summarizing recent views, and re-discussing accepted dogmas on the ram and the buck effect]

Ekström P., Meissl H. (2003). Evolution of photosensory pineal organs in new light: the fate of neuroendocrine photoreceptors. *Philos Trans R Soc Lond B Biol Sci.* 29, 1679–700. [This is very interesting review on pineal gland development and evolution in several classes of vertebrates].

Eldon J. (1993). Effect of exogenous melatonin and exposure to a ram on the time of onset and duration of the breeding season in Icelandic sheep. *Journal of Reproduction and Fertility* 99, 1–6. [This article characterizes the breeding season length in ewes which were permanently exposed to rams]

Evans A.C.O., Duffy P., Crosby T.F., Hawken P.A.R., Boland M.P., Beard A.P. (2004). Effect of ram exposure at the end of progestagen treatment on estrus synchronisation and fertility during the breeding season in ewes. *Animal Reproduction Science* 84, 349–358. [A study on the reproductive response of cyclic ewes to the ram effect]

Fatet A, Pellicer-Rubio MT, Leboeuf B. (2011). Reproductive cycle of goats. *Animal Reproduction Science* 124, 211–219. [This is an up-to-date review of estrous cycles and seasonal reproduction in goats, hormonal and photoperiodic treatments, and buck effect as ways to achieve out-of-season breeding in goats].

Fiol C., Ungerfeld R. (2012). Biostimulation in cattle: stimulation pathways and mechanisms of response. *Tropical and Subtropical Agroecosystems*, in press. [A recent review on the mechanisms by which bulls stimulate prepuberal and postpartum cattle]

Forcada F., Abecia J.A. (2006). The effect of nutrition on the seasonality of reproduction in ewes. *Reproduction Nutrition Development* 46, 355–365. [This is a review of the relationships between nutrition and reproductive seasonality in sheep. It deals with the effects of body fat or food intake on sexual activity in sheep, and the influences of malnutrition and overfeeding].

Giwerzman A., Giwerzman Y.L. (2011). Environmental factors and testicular function. *Best Practice & Research Clinical Endocrinology & Metabolism* 25, 391–402. [This is a review of environmental factors affecting testicular development and function, especially regarding certain environmental chemicals and lifestyle related factors].

Goldman B., Gwinner E., Karsch F.J., Saunders D., Zucker I., Ball G.F. (2004). Circannual rhythms and photoperiodism. In : (Eds : Dunlap J.D., Loros J.J., DeCoursey P.J.), *Chronobiology: biological time keeping*. Sunderland, MA: Sinauer Associates. pp. 107–144. [This is a comprehensive review of circannual rhythms and photoperiodism, studied from a fundamental science biological perspective, in a wide range of animal species].

Gómez-Brunet A., Santiago-Moreno J., del Campo A., Malpoux B., Chemineau P., Tortonese D.J., Gonzalez-Bulnes A., López-Sebastián A. (2008). Endogenous circannual cycles of ovarian activity and changes in prolactin and melatonin secretion in wild and domestic female sheep maintained under a long-day photoperiod. *Biology of Reproduction* 78, 552–562. [This report describes differences in ovulatory activity and endogenous reproductive rhythm between European mouflon (*Ovis orientalis musimon*) wild ewes and Spanish Manchega (*Ovis aries*) domestic ewes under different photoperiodic treatments. This rhythm was differently expressed in the two types of ewe with the domestic animals exhibiting much greater sensitivity to the effects of long days].

Gonzalez R., Orgeur P., Poindron P., Signoret J.P. (1989). Seasonal variation in LH and testosterone responses of rams following the introduction of oestrous ewes. *Animal Reproduction Science* 21, 249–259. [One of the earlier works studying the endocrine responses of rams to estrous ewes]

Gonzalez R., Orgeur P., Poindron P., Signoret J.P. (1991). Female effect in sheep. I. The effects of sexual receptivity of females and the sexual experience of rams. *Reproduction Nutrition Development* 31, 97–102. [Another study on the endocrine response of rams to ewes]

Haim A., Shanas U., Zubidad Ael S., Scantelbury M. (2005). Seasonality and seasons out of time--the thermoregulatory effects of light interference. *Chronobiology International* 22, 59–66. [This is a study performed in wild rodents showing the importance of winter acclimatization of the thermoregulatory system as an example of ambient temperature influences on the expression of seasonal reproduction].

Hawken P.A.R., Beard A.P. (2009). Ram novelty and the duration of ram exposure effects the distribution of mating in ewes exposed to rams during the transition into the breeding season. *Animal Reproduction Science* 111, 249–260. [An experiment in which rams were used before the breeding season to synchronize the ensuing estrous cycles]

Hawken P.A.R., Beard A.P., O'Meara C.M., Duffy P., Quinn K.M., Crosby T.F., Boland M.P., Evans A.C.O. (2005). The effects of ram exposure during progestagen oestrus synchronisation and time of ram introduction post progestagen withdrawal on fertility in ewes. *Theriogenology* 63, 860–871. [An article that presents novel information on the endocrine response of cyclic ewes to the introduction of rams]

Haynes N.B., Haresign W. (1987). Endocrine aspects of reproduction in the ram important to male effect. *World Reviews on Animal Production* XXXIII, 21–28. [A review on rams characteristics and management which can be of help to maximize the response of ewes to the ram effect]

Knight T.W. (1985). Are rams necessary for the stimulation of anoestrous ewes with oestrous ewes? *Proceedings of the New Zealand Society of Animal Production* 45, 49–50. [This work aimed to stimulate anoestrous ewes with estrous ewes, but at the same time determined the advantages of including estrous ewes during the ram effect stimulation.]

Kumar V. (1997). Photoperiodism in higher vertebrates: an adaptive strategy in temporal environment. *Indian Journal of Experimental Biology* 35, 427–437. [A review on the photoperiodism and the mechanisms transmitting photic information from the eyes to the pineal gland and the endocrine organs].

Lincoln G.A., Clarke IJ, Hut RA, Hazlerigg D.G. (2006). Characterizing a mammalian circannual pacemaker. *Science*. 314, 1941–4. [A review on the anatomy and physiology of the internal circannual rhythm regulation in sheep].

Lindsay D.R., Gray S.J., Oldham C.M., Pearce D.T. (1984). The single injection of progesterone. *Proceedings of the Australian Society of Animal Production* 15, 159–161. [The study describes that a single administration of progesterone to anoestrous ewes at the time in which rams are introduced hampers short lived corpora lutea and synchronizes estrous behavior]

Lindsay D.R., Wilkins J.F., Oldham, C.M. (1992). Overcoming constraints: the ram effect. *Proceedings of the Australian Society of Animal Production* 19, 208–210. [A short view on practical applications of the ram effect]

Lucidi P., Barboni B., Mattioli M. (2001). Ram-induced ovulation to improve artificial insemination efficiency with frozen semen in sheep. *Theriogenology* 55, 1797–1805. [The ram effect has been used in cyclic ewes to improve the fertility obtained in inseminated ewes]

Macchi E., Cucuzza A.S., Badino P., Odore R., Re F., Bevilacqua L., Malfatti A. (2010). Seasonality of reproduction in wild boar (*Sus scrofa*) assessed by fecal and plasmatic steroids. *Theriogenology* 73, 1230–1237. [A study on the reproductive seasonality of two wild boars populations, living in a mountainous vs. a plain habitat in Italy].

Martin G.B., Oldham C.M., Cognié Y., Pearce D.T. (1986). The physiological responses of anovulatory ewes to the introduction of rams - a review. *Livestock Production Science* 15, 219–247. [A classical review summarizing knowledge on the ram effect]

Martin G.B., Walkden-Brown S.W. (1995). Nutritional influences on reproduction in mature male sheep and goats. *Journal of Reproduction and Fertility, Suppl.* 49, 437–449. [A relevant review summarizing the nutritional influences on reproductive seasonality of male sheep and goats].

Mauget R. (1972). Observations on reproduction in wild swine (*Sus scrofa* L.). *Annales de Biologie Animale, Biochimie, Biophysique*. 12, 195–202. [A pioneer study on the reproductive seasonality of wild boars].

Meikle A., Forsberg M., Garófalo E.G., Carlsson M.A., Lundeheim N. Rubianes E. (2001). Circulating gonadotrophins and follicular dynamics in anoestrous ewes after treatment with estradiol-17beta. *Animal Reproduction Science* 67, 79–90. [This is the first experiment in which it was demonstrated that estradiol can be used to synchronize the emergence of follicular waves in ewes]

Mossman A.S., Mossman H.W. (1962). Ovulation, implantation, and fetal sex ratio in impala. *Science* 14, 869. [A study on the reproductive biology of African savanna's wild impalas].

Muir P.D., Smith N.B., Wallace G.J. (1989). Early lambing in Hawkes Bay: use of the ram effect. *Proceedings of the New Zealand Society of Animal Production* 49, 271–275. [A practical view on the use of the ram effect]

Ngere L.O., Dzakuma J.M. (1975). The effect of sudden introduction of rams on oestrus pattern of tropical ewes. *Journal of agricultural Sciences (Cambridge)* 84, 263–264. [One of the earlier reports on the reproductive response of cyclic ewes to the ram effect]

Nugent R.A. III, Notter D.R. (1990). Effects of cohabitation with white-faced ewes on estrous activity of Hampshire and Suffolk ewes exposed to rams in June. *Journal of Animal Science* 68, 1513–1519. [The influence of anestrus depth on the response to the ram effect was studied in this paper]

O’Callaghan D., Donovan A., Sunderland S.J., Boland M.P., Roche J.F. (1994). Effect of the presence of male and female flockmates on reproductive activity in ewes. *Journal of Reproduction and Fertility* 100, 497–503. [In this study several experiments were included to demonstrate the influence of the continuous presence of rams or other ewes on the breeding season length]

Ortavant R., Bocquier F., Pelletier J., Ravault J.P., Thimonier J., Volland-Nail P. (1988). Seasonality of reproduction in sheep and its control by photoperiod. *Australian Journal of Biological Sciences* 41, 69–85. [This review discusses data backing up the hypothesis on photoperiodic time measurement in sheep].

Pearce D.T., Martin G.B., Oldham C.M. (1985). Corpora lutea with a short life-span induced by rams in seasonally anovulatory ewes are prevented by progesterone delaying the preovulatory surge of LH. *Journal of Reproduction and Fertility* 84, 333–339. [In this study it was proposed that a single injection of progesterone prevents the occurrence of short-lived corpora lutea. Although it was proposed that the mechanism was a delay in LH surge, later it was demonstrated by the same group that this was not the mechanism involved. In any case, it is a clear report on the endocrine and ovarian response of anestrus ewes to the ram effect]

Pearce D.T., Oldham C.M. (1983). Ram effect in the breeding season. *Proceedings of the Australian Society of Reproductive Biology*, 4-7 September, Canberra, Australia, p. 49. [An abstract reporting that there are endocrine responses of cyclic ewes to the ram effect]

Pearce D.T., Oldham C.M. (1988). Importance of non-olfactory ram stimuli in mediating ram-induced ovulation in the ewe. *Journal of Reproduction and Fertility* 84, 333–339. [This is a pioneer study that aimed to determine the importance of different signals of rams in the response of anestrus ewes]

Prendergast B. (2005). Internalization of seasonal time. *Hormones and Behavior* 48, 503–511. [This review discusses photorefractoriness and reproductive activity recrudescence in wild rodents].

Ravault J.P., Martinat-Botte F., Mauget R., Martinat N., Locatelli A., Bariteau F. (1982). Influence of the duration of daylight on prolactin secretion in the pig: hourly rhythm in ovariectomized females, monthly variation in domestic (male and female) and wild strains during the year. *Biology of Reproduction* 27, 1084–1089. [This review discusses reproductive seasonality patterns in both domestic and wild pigs].

Rensing L., Meyer-Grahe U., Ruoff P. (2001). Biological timing and the clock metaphor: oscillatory and hourglass mechanisms. *Chronobiology International* 18, 329–369. [A review discussing the mechanisms of biological timing in different organisms].

Rodríguez Iglesias R.M., Ciccio N., Irazoqui H. (1997). Ram-induced reproduction in seasonally anovular Corriedale ewes: MAP doses for oestrus induction, ram percentages and postmating progestagen supplementation. *Animal Science* 64, 119–125. [An article in which some mechanisms to improve the response to the ram effect were studied. In relation to progestagen primings, in this study the influence of the amount of medroxyprogesterone contained in the intravaginal sponges was determined]

Rodríguez Iglesias R.M., Ciccio N., Irazoqui H., Rodríguez B.T. (1991). Importance of behavioural stimuli in ram-induced ovulation in seasonally anovular Corriedale ewes. *Applied Animal Behaviour Science* 30, 323–332. [An elegant study on how the different signals provided by rams and estrus ewes interact to improve the response of anestrus ewes to the ram effect]

Rosa H.J.D., Bryant M.J. (2002). Review: The “ram effect” as a way of modifying the reproductive activity in the ewe. *Small Ruminant Research* 45, 1–16. [A review summarizing knowledge on the ram effect]

Rubianes E., de Castro T., Carbajal B. (1996). Effect of high progesterone levels during the growing phase of the dominant follicle of wave 1 in ultrasonically monitored ewes. *Canadian Journal of Animal Science* 76, 473–475. [In this article the effects of high progesterone concentrations on follicular dynamics in ewes was described]

Sadleir RM. (1968). Reproductive responses to the environment in mammals. *Journal of Psychosomatic Research* 12, 3–9. [A review discussing the main environmental factors which influence reproductive activity in mammals].

Scaramuzzi RJ, Martin GB. (2008). The importance of interactions among nutrition, seasonality and socio-sexual factors in the development of hormone-free methods for controlling fertility. *Reproduction in Domestic Animals* 43 Suppl 2, 129-136. Interesting review of possible applications of environmental factors as hormone-free methods for fertility control in ruminants.

Schlatt S., De Geyter M., Kliesch S., Nieschlag E., Bergmann M. (1995). Spontaneous recrudescence of spermatogenesis in the photoinhibited male Djungarian hamster, *Phodopus sungorus*, *Biology of Reproduction* 53, 1169–1177. [A review on the reproductive seasonality, and internal circannual rhythms of male hamsters].

Simpson S.M., Follett B.K., Ellis D.H. (1982). Modulation by photoperiod of gonadotrophin secretion in intact and castrated Djungarian hamsters. *Journal of Reproduction and Fertility* 66, 243–250. [A review on photoperiodic influences acting on the hormones regulating reproductive activity in hamsters].

Smith H.J., McLaren J.B., Odom J.A., Miller H. (1958). Influence of the use of sterile teaser rams prior to breeding on subsequent fertility of ewes. *Journal of Animal Science* 17, 1231 (abstr). [An abstract in which infertile rams were used before introducing a fertile ram in order to synchronize estrous cycles]

Spencer T.E., Becker W.C., George P., Mirando M.A., Ogle T.F., Bazer F.W. (1995). Ovine interferon-tau inhibits estrogen receptor up-regulation and estrogen-induced luteolysis in cyclic ewes. *Endocrinology* 136, 4932–4944. [A study clarifying luteolytic mechanisms in sheep]

Stumpf T.T., Wolfe M.W., Roberson M.S., Kittok R.J., Kinder J.E. (1993). Season of the year influences concentration and pattern of gonadotropins and testosterone in circulation of the bovine male. *Biology of Reproduction* 49, 1089–1095. [This study describes mild seasonal variations in reproductive hormones secretion in domestic bulls].

Taberlet P., Coissac E., Pansu J., Pompanon F. (2011). Conservation genetics of cattle, sheep, and goats. *Comptes rendus biologiques* 334, 247–254. [This review deals with self sustaining management strategies of biodiversity in domestic ruminants].

Tast A., Peltoniemi O.A., Virolainen J.V., Love R.J. (2002). Early disruption of pregnancy as a manifestation of seasonal infertility in pigs. *Animal Reproduction Science* 74, 75–86. An article describing fertility problems associated to seasonal reproductive patterns in pigs.

Ungerfeld R. (2006). Socio-sexual signalling and gonadal function: Opportunities for reproductive management in domestic ruminants. En: Juengel, J.I., Murray, J.F., Smith, M.F. (Eds.), *Reproduction in Domestic Ruminants VI*, Nottingham University Press, Nottingham, UK, pp. 207–221. [A review on how social stimuli can be used to stimulate non-cyclic ewes, does, and cows]

Ungerfeld R. (2008). Response of anestrus ewes pretreated with a single dose of oestradiol-17beta or progesterone and oestradiol-17beta to the introduction of rams and ewes in oestrus. *New Zealand Veterinary Journal* 56, 36–39. [In this study it was observed that as estradiol alters the follicular development in anestrus ewes, and the ovarian response to the ram effect is affected by estradiol administration, the ovarian response to the ram effect was related to the follicular development at the time that rams are introduced]

Ungerfeld R. (2009). The induction of oestrus in ewes during the non-breeding season using pre-used CIDRs and oestradiol-17beta treatment. *Small Ruminant Research* 84, 129–131. [As intravaginal devices containing progesterone – as CIDRs– can be reused, in this experiment the aim was to improve the reproductive response of ewes primed with previously used CIDRs with estradiol administration]

Ungerfeld R. (2011). Combination of the ram effect with PGF2 α estrous synchronization treatments in ewes during the breeding season. *Animal Reproduction Science* 124, 65–68. [In this article it was demonstrated that the stimulation with rams improves estrus synchronization results in cyclic ewes. However, a complementary hypothesis i.e., the possible luteolytic effect of ram introduction was not demonstrated]

Ungerfeld R., Carbajal B., Rubianes E., Forsberg M. (2005). Endocrine and ovarian changes in response to the ram effect in medroxy-progesterone acetate-primed Corriedale ewes during the breeding and the

nonbreeding season. *Acta Veterinaria Scandinavica* 46, 33–44. [A study on the endocrine and ovarian responses of medroxyprogesterone primed ewes to the ram effect]

Ungerfeld R., Dago A.L., Rubianes E., Forsberg M. (2004). Response of anestrus ewes to the ram effect after follicular wave synchronization with a single dose of estradiol-17beta. *Reproduction Nutrition Development* 44, 89–98. [The relationship of follicular growth status and the ovarian response pattern to the ram effect was studied. Follicular waves were synchronized with estradiol, but there was not a clear link between follicular status and response]

Ungerfeld R., Forsberg M., Rubianes E. (2004). Overview of the response of anoestrous ewes to the ram effect. *Reproduction Fertility Development* 16, 479–490. [An updating review on the ram effect]

Ungerfeld R., Pinczak A., Forsberg M., Rubianes E. (2002). Ovarian and endocrine responses of Corriedale ewes to "ram effect" in the non-breeding season. *Canadian Journal of Animal Science* 82, 599–602. [The ovarian response of anoestrous ewes was not so simple as was previously described. Using ultrasound the authors described ovarian responses not previously observed]

Ungerfeld R., Pinczak A., Forsberg M., Rubianes E. (1999). Response of Corriedale ewes to the "ram effect" after primings with medroxyprogesterone, fluorogestone, or progesterone in the non-breeding season. *Acta Veterinaria Scandinavica* 40, 299–305. [In this study it was demonstrated that any of the progestagens tested as primings to the ram effect provides the same results]

Ungerfeld R., Ramos M.A., González-Pensado S.P. (2008). Ram effect: adult rams induce a greater reproductive response in anoestrous ewes than yearling rams. *Animal Reproduction Science* 103, 271–277. [Although some authors proposed that adult rams are more effective than young rams to stimulate anoestrous ewes, this was the first study demonstrating this. In this study it was also demonstrated that chemical signals from adult rams are responsible for this difference]

Ungerfeld R., Rubianes E. (1999a). Effectiveness of short-term progestogen priming for the induction of fertile oestrus with eCG in ewes during late seasonal anoestrus. *Animal Science* 68, 349–353. [Although progestagen primings are usually used 12-14 days in ewes, this study demonstrated that at least in anoestrous ewes 6 days of application are enough to obtain maximum responses]

Ungerfeld R., Rubianes E. (1999b). Estrous response to ram effect in Corriedale ewes primed with medroxyprogesterone during the breeding season. *Small Ruminant Research* 32, 89–91. [A simple study demonstrating that the ram effect improves the response of cyclic ewes to progestagen based estrous synchronization treatments]

Ungerfeld R., Suárez G., Carbajal B., Silva L., Laca M., Forsberg M., Rubianes, E. (2003). Medroxyprogesterone primings and response to the ram effect in Corriedale ewes during the non-breeding season. *Theriogenology* 60, 35–45. [Three experiments were performed to determine minimum times and amounts for medroxyprogesterone primings application in combination with the ram effect]

Yarney T.A., Sanford L.M. (1983). The reproductive-endocrine response of adult rams to sexual encounters with estrual ewes is season dependent. *Hormones and Behavior* 17, 169–182. [The differences on the endocrine response of rams to estrous ewes during the breeding and the non breeding season were studied]

Whitten W.K. (1956). Modification of the oestrous cycle of the mouse by external stimuli associated with the male. *Journal of Endocrinology* 13, 399–404. [The first study demonstrating that male stimulation may synchronize estrous cycles in mammals]

Zarazaga L.A., Guzmán J.L., Domínguez C., Pérez M.C., Prieto R. (2005). Effect of plane of nutrition on seasonality of reproduction in Spanish Payoya goats. *Animal Reproduction Science* 87, 253–267. [This study reports the effect of the plane of nutrition on the reproductive seasonality of Mediterranean origin goats].

Zuluaga J.F., Williams G.L. (2008). High-pressure steam sterilization of previously used CIDR inserts enhances the magnitude of the acute increase in circulating progesterone after insertion in cows. *Animal Reproduction Science* 107, 30–35. [Previously used CIDRs were used again in cows, and the progesterone serum concentrations achieved were similar than those obtained with new CIDRs if used CIDRs were autoclaved]

Biographical Sketches

Rodolfo Ungerfeld, Biologist (Universidad de la República, Uruguay), Master in Biology-Physiology (Programa de Desarrollo de Ciencias Básicas, PEDECIBA, Uruguay), PhD (Swedish University of Agricultural Sciences, Sweden). Full Professor of Animal Physiology at the Facultad de Veterinaria (Universidad de la República, Uruguay). Author of more than 70 peer-reviewed articles in the field of reproduction, endocrinology and behavior in ruminants, including work with sheep, goats, cattle, and deer. Scientific reviewer for more than 30 international journals, and member of the Editorial Committee of 3 journals. Researcher of PEDECIBA, member of the standing Committee of Postgraduate Studies. Winner of the Prize of the National Academy of Veterinary Science, Uruguay (1997, 2004, and 2009), and of the Caldeyro-Barcia Award (most important scientific award in Uruguay) in 2003.

Alejandro Bielli, graduated in Veterinary Medicine and Technology at the Universidad de la República (Montevideo, Uruguay) in 1992 and took graduate studies (Licentiate in Veterinary Medicine, 1995 and PhD, 1999, both in histophysiology of seasonal testicular activity of rams, and the interaction with nutrition) at the Swedish University of Agricultural Sciences (Uppsala, Sweden). Full Professor at the Facultad de Veterinaria, Universidad de la República (Uruguay). Research interests: nutritional influences on seasonal reproduction, fetal programming and early life nutritional influences on reproductive activity. Author of 25 research papers in international journals, 4 chapter books and numerous presentations in international scientific meetings on animal reproduction and histology. Researcher of PEDECIBA, Uruguay (2003), Member of the standing Committee of Graduate Studies, Veterinary Graduate Studies Program, University of Uruguay (2006-), winner of the Prize of the National Academy of Veterinary Science, Uruguay (1999 and 2010).